# Current Status on Single Cell Protein (SCP) Production from Photosynthetic Purple Non Sulphur Bacteria

Swetha Garimella<sup>1</sup>, Karunakar Rao Kudle<sup>2</sup>, Aruna Kasoju<sup>3</sup>, Ramchander Merugu<sup>1</sup>\*

<sup>1</sup>University College of Science and Informatics, Mahatma Gandhi University, Nalgonda, India-508 254

<sup>2</sup>Department of Biochemistry, Osmania University, Hyderabad-500007, India-500 007

<sup>3</sup>Department of Biotechnology, Jawaharlal Nehru Technological University, Anantapur, India-515 002

\*Corresponding author: E-Mail: rajumerugu01@gmail.com ABSTRACT

Bacterial production of SCP have a significant advantage over other microorganisms. Single cell protein is the total protein extracted from the bacterial cultures. Apart from the nutritional benefits of single cell protein, single cell protein technology is sustainable technology. Prototroph's have high protein content with essential amino acids and less amounts of nucleic acids. PNSB are highly resistant to toxicants and contain 70-72% of crude protein. The composition of essential amino acids of these bacteria is similar to that of soyabean proteins. The application of PNSB has been used to convert to hydrogen gas (Liao, 2010) or by products such as single cell proteins. PNSB biomass is useful for fish feeding as it is rich in proteins. An in-depth analysis of the single cell production abilities of PNSB show that the purple non sulphur bacterium Rhodopseudomonas palustris would be best in terms of single cell protein production.

KEY WORDS: PNSB, Single cell protein (SCP), Rhodopseudomonas, Rhodobacter, Bioremediation.

#### 1. INTRODUCTION

Advances in present day biology in Single cell protein (SCP) production processes have led to the use of microorganism rather than animal or plant sources. Recycling of waste by bacteria to protein production is both economical and environmental friendly. Bacterial production of SCP have a significant advantage over other microorganisms. Single cell protein is the total protein extracted from the bacterial cultures. Apart from the nutritional benefits (Adedayo, 2011) of single cell protein, single cell protein technology is sustainable technology. SCP refers to total proteins extracted from pure microbial cell culture and is produced using microorganisms including bacteria, fungus and algae (Anupama & Ravindra, 2000). The use of natural cheap substrates is explored for cultivating microorganisms (Grewal, 1990; Osho, 1995). Microorganisms have a much higher protein content of 30-70% in the dry mass and in particular PNSB have more than 50% protein content. Single cell protein aminoacid composition of microbial SCP has good nutritional properties (Adedayo, 2011). Generation time of Methylophilus is about 2 hours which saves time and large amounts of biomass can be generated. Large amounts of SCP are produced by bacteria like Acromobacter delvaevate, Acinetobacter calcoacenticus, Aeromonas hydrophilla, Brevibacterium (Adedayo, 2011), Bacillus subtilis (Gomashe, 2014), Flavobacterium sps, Lactobacillus sps, Cellulomonas species, Methylophilus methylitropous (Jenkins, 1987), Pseudomonas fluorescens, Thermomonospora fusca (Dhanasekaran, 2011). Single Cell Protein (SCP) is the best way of solving global food problem (Najafpour, 2007). If waste materials are used as substrate, waste management can also be investigated. Proteins not only function as nutrients but also perform other functions (Mahajan and Dua, 1995; Asad, 2000; Jamel 2008; Nasseri, 2011). Hydrocarbons, polysaccharides and other wastes are used for SCP production (Ashok, 2000 & Atalo and Gashe 1993). Microbial protein productions with several natural products such as grape juice byproduct, date extract, cashew apple juice have been used for production of SCP (Kuzmanova, 1989). Cellulsoe was also investigated for SCP production (Bozakuk, 2002; Zubi, 2005).

Bacterial SCP production has advantages such as rapid growth of bacteria and short generation time. They are also capable of growing on starch, sugars, methane, petroleum fractions, alcohols and various nitrogen sources including ammonia (Bamberg, 2000). The substrates which are in use are gaseous hydrocarbons, liquid hydrocarbons and alcohols are utilized by bacteria. Methanogenic bacteria Istrains are recommended for SCP production (Arora, 1991). A number of industrial waste products are being used for producing SCP (Smith and Bull, 1976; Saquido, 1981; Abou Hamed, 1993; Tipparat, 1995; Nigam, 2000; Zhao, 2010).

PNSB for SCP Production: Usage of photosynthetic bacteria for bioremediation is being explored as microorganisms are capable of removing organic substances in waste water (Hiraishi, 1989; Madigan, 2000; Noparatnaraporn, 1986) producing byproducts such as protein, carotenoids, hopanoids, phytohormones, aminolevulinic acid and vitamins (Kobayashi and Tchan, 1973; Getha, 1998; Banerjee, 2000). They have flexible metabolic pathways and being used for wastewater purification in recent times (Siefert, 1978; Howard, 1987; Imhoff, 1992; Kantachote, 2005; Yegani, 2005; Chae, 2006). Purple non sulphur photosynthetic bacteria (PNSB) are ) are versatile organisms which widely distributed in different ecological niches (Holt, 1994; Zhu, 2002; Hoogewerf, 2003; Holt, 1994; Okubo, 2006; Kaewsuk, 2010). Anupama and Ravindra, (2000) suggested bacterial production of SCP has more advantages than other groups. Prototroph's have high protein content with essential amino acids and less

## Journal of Chemical and Pharmaceutical Sciences

amounts of nucleic acids (Sasikala and Ramana, 1995, Merugu, 2008). PNSB are highly resistant to toxicants (Ding, 2008; Madukasi, 2010) and contain 70-72% of crude protein (Kim and Lee, 2000). Kim and Lee (2000), studied the use of Rhodopseudomonas palustris as fish feed. The composition of essential amino acids of these bacteria is similar to that of soyabean proteins (Azad, 2001). The application of PNSB has been used to convert to hydrogen gas (Liao, 2010) or by products such as single cell proteins. PNSB biomass is useful for fish feeding as it is rich in proteins (Honda, 2006). The factors which can be optimized for PNSB cultivation are anaerobic condition, light (in lux), pH, carbon, nitrogen, agitation and growth factors (Honda, 2006; Kaewsuk, 2010). Growth has been studied in photosynthetic pond (Honda, 2006), batch photo-bioreactor and membrane photo-bioreactor (Kaewsuk, 2010). Methionine and phenylalanine concentrations were low (Noparatnaraporn and Nagai, 1986) in Rubrivivax (Rhodocylcus) gelatinosus R1 biomass when compared to other similar findings of other authors. R. gelatinosus R1 biomass aminoacid composition was same that of algae (Prasertsan, 1993). Essential fatty acids are essential for growth of fish (Izquierdo, 1996) which are present in these bacterial feed (Loo, 2012). Kim and Lee (2000), estimated USD 79 per kg production of PNSB but this can be made cost effective if agro industrial wastewaters are used as substrate. Merugu, (2012) has reported photosynthetic bacterial growth in cellulose has carbon source. Bender & Phillips (2004) and Banerjee, (2000) suggested the use of PNSB as cheaper alternative protein sources for fish feed. Rhodocyclus gelatinosus found in many industrial effluents hydrolyzes starch and gelatin (Holt, 1994) and may find application in enhancing their colour due to production of spirilloxanthin series of pigments (Kobayashi and Kurata, 1978; Noparatnaraporn and Nagai, 1986). Salma, (2007) has recommended purple non sulphur bacteria as a source of single cell protein. Imhoff (2006) suggested the use of PNSB with simultaneous SCP production. Kantachote, (2005) reported a protein percentage of 66.7 from Wastewater from a latex rubber sheet processing from Rhodopseudomonas blastica. About 72-74% of SCP protein from sludge and sago starch processing using Rhodopseudomonas palustris was observed by Geetha, (1998) which was used as aquaculture feed. Elisa, (2002) observed 67.6% of SCP from Rhodocyclus gelatinosus cultured in Poultry slaughterhouse wastewater which was used as feed supplement. Napavarn & Nagai (1986), used Rhodobacter sphaeroides P47 cultured in Pineapple waste and reported 6.6% SCP. Rhodopseudomonas palustris P1cultured in fermented pineapple extract contained 65% SCP and used for the treatment of latex rubber sheet wastewater (Nastee, 2014). He, (2010) reported 52% SCP from Rhodobacter sphaeroides Z08 grown in Soybean waste water. Rhodovulvum sulphidophilum grown in Glutamate Malate medium produced 15.6% SCP (Maneewan, 2010). Elisa (2003), reported 50.6% from Rhodocyclus gelatinosus cultivated from pig farm waste. Balloni, (1987) reported 58% SCP from Rhodopseudomonas and R. fulvum cultivated from sugar refinery wastewater. Rhodocyclus gelatinosus grown in miso-like effluent medium produced 63% SCP (Sasaki, 1981). Rhodobacter sphaeroides P47 grown in dehydrated medium from Pineapple peel waste produced 66.6% SCP (Noparatnaraporn, & Nagai, 1986). Prasertsan, (1993) reported 50% SCP from Rhodocyclus gelatinosus cultivated in seafood processing wastewater. Rhodopseudomonas sp. grown in Synthetic 112 medium produced 11% SCP (Tan, 2014). Rhodopseudomonas palustris & Rhodobacter blasticus grown in wastewater from noodle production produced 50% SCP (Worowit, 2016). Saejung & Thammaratana (2016) reported 60.1% SCP production from culturing Rhodopseudomonas sp. CSK01 cultured from Municipal wastewater. Rhodocyclus gelatinosus R7 produced 56% SCP from Tuna condensate (Poonsuk, 1997). Alexandre, (2009) reported 45% SCP from Rhodobacter capsulatus cultivated from Synthetic medium. Rhodopseudomonas acidophila produced 23% SCP from synthetic medium (Pfennig, 1969). Napavarn, (1987) reported 56% SCP from Rhodocyclus gelatinosus cultivated in Cassava waste. Rhodopseudomonas palustris grown in simulated waste water produced 45% SCP (Honda, 2006). Kim, (1999) reported 74% SCP Rhodopseudomona spalustris grown in photosynthetic sludge which was used as diet for aquaculture. Rhodopseudomonas capsulatus grown in cow dung contained essential sulphur amino acids (Gupta, 2016). Rhodobacter sphaeroides grown in Cassava waste also produced essential aminoacids which was used as Fish meal (Napavarn, 1986).

Table.1. SCP Production from Photosynthetic Purple Non Sulphur Bacteria

Organisms	Source of organism	Protein (%, mg/g dry wt)	Application	Reference
Rhodopseudomon as blastica	Wastewater from a latex rubber sheet processing	66.7	Proposed to be used as Animal feed	Kantachote, (2005)
Rhodopseudomon as palustris	Sludge and sago starch processing	72–74	As aquaculture feed	Geetha, (1998)
Rhodocyclus gelatinosus	Poultry slaughterhouse wastewater	67.6	Poultry feed	Elisa, (2002)
Rhodobacter sphaeroides P47	Pineapple waste	66.6	Feed supplement	Napavarn & Nagai (1986)

ISSN: 0974-2115

# www.jchps.com

# Journal of Chemical and Pharmaceutical Sciences

www.jchps.com		Journal of Chemical and Pharmaceutical Sciences			
Rhodopseudomon as palustris P1	Fermented pineapple extract	65	Treatment of latex rubber sheet wastewater	Nastee (2014)	
Rhodobacter sphaeroides Z08	Soyabean waste water	52	Bioremediation	He, (2010)	
Rhodovulvum sulphidophilum	Glutamate Malate Medium	15.6	Fish feed supplement	Maneewan (2010)	
Rhodocyclus gelatinosus	Pig farm waste	50.6	Remediation	Elisa, (2003)	
Rhodopseudomon as and R. fulvum	Sugar refinery wastewater	58	Remediation	Balloni, (1987)	
Rhodocyclus gelatinosus	miso-like effluent medium	63	Remediation	Sasaki, (1981)	
Rhodobacter sphaeroides P47	Dehydrated medium from Pineapple peel waste	66.6	Remediation	Noparatnarapor n, & Nagai (1986)	
Rhodocyclus gelatinosus	Seafood processing wastewater	50	Remediation	Prasertsan, (1993)	
Rhodopseudomon as sp.	Synthetic 112 medium	11	Remediation	Tan, (2014)	
Rhodopseudomon as palustris & Rhodobacter blasticus	Wastewater from noodle production	50	Remediation	Worowit, (2016)	
Rhodopseudomon as sp. CSK01	Municipal wastewater	60.1	Remediation	Saejung & Thammaratana (2016)	
Rhodocyclus gelatinosus R7	Tuna condensate	56	Remediation	Poonsuk, (1997)	
Rhodobacter capsulatus	Synthetic medium	45	Remediation	Alexandre, (2009)	
Rhodopseudomon as acidophila	Synthetic medium	23	Remediation	Pfennig (1969)	
Rhodocyclus gelatinosus	Cassava waste	56	Remediation	Napavarn, (1987)	
Rhodopseudomon as palustris	Simulated waste water	45	Remediation	Honda, (2006)	
Rhodopseudomon aspalustris	Photosynthetic sludge	74	Diet for aquaculture	Kim, (1999)	
Rhodopseudomon as capsulatus	Cow dung	Essential sulphur amino acids	Fish feed	Gupta, (2016)	
Rhodobacter sphaeroides	Cassava waste	Essential aminoacids	Fish meal	Napavarn, (1986)	

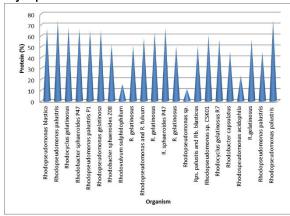


Figure.1. Graphical Representation Of Scp Production By Pnsb

## Journal of Chemical and Pharmaceutical Sciences

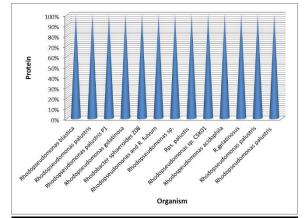


Figure.2. Graphical representation Of SCP production by Rhodopseudomonas genus

#### 2. CONCLUSION

An in-depth analysis of the single cell production abilities of PNSB show that the purple non sulphur bacterium *Rhodopseudomonas palustris* would be best in terms of single cell protein production. Further work for SCP production should involve using *Rhodopseudomonas palustris* instead of other genus in this group as this organism is more versatile and best suited for the production of Single cell protein.

#### REFERENCES

Abou Hamed S.A.A, Bioconversion of wheat straw by yeast into single cell protein, Egypt. J. Microbiol, 28 (1), 2013, 1-9.

Adedayo M.R, Ajiboye E.A, Akintunde J.K, Odaibo A, Single Cell Proteins, As Nutritional Enhancer, Advances in Applied Science Research, 2 (5), 2011, 396-409.

Alexandre J, Poulain and Dianne K, Newman, *Rhodobacter capsulatus* catalyzes Light-Dependent Fe (II) Oxidation under Anaerobic Conditions as a Potential Detoxification Mechanism. Applied and Environmental Microbiology, 2009, 6639–6646.

Anupama and Ravindra, Value added Food, single cell protein, Biotechnol. Adv. J. Microbiol, 18, 2000, 459 479.

Arora D, Mukerji K & Marth E, Single cell protein in Hand book of applied mycology, India, Banaras Hindu University, 3, 1991, 499-539.

Asad M.J, Asghan M, Yaqub M and Shahzad K, Production of single cell protein delignified corn cob by Arachniotus species, Pak. J. of Agric. Sci, 37, 2000, 3-4.

Ashok Pandey, Carlos Soccol R, Poonam Nigam, Vanete Soccol T, Luciana P.S, Vandenberghe, Radjiskumar Mohan, Biotechnological potential of agro-industrial residues. II: cassava bagasse, Bio resource Technology, 74, 2000, 81-87

Atalo K and Gashe BA, Protease production by a thermophilic Bacillus species (P-001 A) which degrades various kinds of fibrous proteins, Biotechnology Letters, 15, 1993, 1151-1156.

Azad S.A, Chong V.C, Vikineswary S & Ramachandran K.B, Potential of waste grown phototrophic bacteria as a feed ingredient in aqua feeds, J. Biosci, 12, 2001, 1–11.

Balloni W, Carlozzi P, Ventura S, De Philippis R and Bosco M, A three years experiment on the production of *Rhodopseudomonas* and *Rhodospirillum* biomass by outdoor culture on different wastes. 4<sup>th</sup> Int. Conference on Biomass for Energy and Industry, Orleans, 1987.

Bamberg J.H, British Petroleum and global oil, 1950-1975, the challenge of nationalism, Volume 3 of British Petroleum and Global Oil 1950-1975, The Challenge of Nationalism, J.H. Bamberg British Petroleum series. Cambridge University Press, 3, 2000, 426–428.

Banerjee S, Azad S.A, Vikineswary S, Selvaraj O.S & Mukherjee T.K, Phototrophic bacteria as fish feed supplement. Asian-Australian Journal of Animal Science 13, 2000, 991-994

Bender J and Phillips P, Microbial mats for multiple applications in aquaculture and bioremediation. Bioresour Technol, 94, 2004, 229–238

Benemann J.R, Microalgae Aquaculture Feeds, J. Applied Phycol, 4, 1992, 233-245.

## Journal of Chemical and Pharmaceutical Sciences

Chae S.A, Hwang F, Akira H, Single cell protein production of Euglena gracilis and carbondioxide fixation in an innovative photobioreactor, Bioresour. Tech, 97 (2), 2006, 322-329.

Ding C, Experiments on the biodegradation of phenol wastewater by immobilized photosynthetic bacteria, Water Resour. Prot, 24 (3), 2008, 93-95.

Elisa Helena Giglio Ponsano, Pedro Magalhães Lacava, Marcos Franke Pinto Chemical composition of *Rhodocyclus gelatinosus* biomass produced in poultry slaughterhouse wastewater, Braz. Arch. Biol. Technol, 46 (2), 2003.

Gabriel A, Ntuli V, James D, Cactus pear biomass, a potential lignocellulose raw material for Single Cell Protein production (SCP), Int. J. Curr. Microbiol. Appl. Sci, 3 (7), 2014, 171-197.

Getha K, Vikineswary S and Chong V.C, Isolation and growth of the phototrophic bacterium Rhodopseudomonas palustris Strain B1 in sago-starch processing wastewater. World Journal of Microbiology and Biotechnology, 14, 1998, 505–511.

Gomashe Ashok V, Pounikar Minakshi A and Gulhane Pranita A, Liquid Whey, A Potential Substrate for Single Cell Protein Production from Bacillus subtilis NCIM 2010. Int. J. of Life Sciences, 2 (2), 2014, 119-123.

Hiraishi A, Shi J.L and Kitamura H, Effects of organic nutrient strength on the purple non-sulfur bacterial content and metabolic activity of photosynthetic sludge for wastewater treatment, J. Ferment. Bioeng, 68 (4), 1989, 269-276.

Holt J.G, Krieg N.R, Sneath P.H.A, Staley J.T and Williams S.T, Bergey's Manual of Determinative Bacteriology, Williams & Wilkins, Baltimore, 9, 1994, 787.

Honda R, Fukushi K, Yamamoto K, Optimization of wastewater feeding for single-cell protein production in an anaerobic wastewater treatment process utilizing purple non-sulfur bacteria in mixed culture condition, J. Biotechnology, 125, 2006, 565-573.

Hoogewerf G.J, Jung D.O and Madigan T, Evidence for limited species diversity of bacteriochlorophyll *b*-containing purple nonsulfur anoxygenic phototrophs in freshwater habitats, FEMS Microbiology Letters, 218 (2), 2003, 359-364.

Howard G, The world of microbes, The Benjamin Cummings publishing, Inc, 1987.

Imhoff J.F, Taxonomy, phylogeny and general ecology of anoxygenic phototrophic bacteria. Mann. N.H, Carr N.G, (Eds.), Bergey's manual of determinative bacteriology, Plenum press New York, 1992.

Imhoff J.F, The phototrophic alphaproteobacteria. In The Prokaryotes, A Handbook on the Biology of Bacteria. Proteobacteria, Alpha and Beta Subclass, edited by Dworkin M, Falkow S & Rosenberg E, New York, Springer-Verlag, 2006, 41-64.

Izquierdo M.S, Essential fatty acid requirements of cultured marine fish larvae, Aquacult. Nutr, 2, 1996, 183-191.

Izu K, Nakajima F, Yamamoto K and Kurisu F, Aeration conditions affecting growth of purple nonsulfur bacteria in an organic wastewater treatment process. Systematic and Applied Microbiology, 24 (2), 2001, 294-302.

Jamel P, Alam M.Z and Umi N, Media optimization for bio proteins production from cheaper carbon source, J. of Engi. Sci. and Techno, 3 (2), 2008, 124-130.

Joong Kyun Kim, Bum-Kyu Lee, Sang-Hee Kim, Jung-Hye Moon, Characterization of denitrifying photosynthetic bacteria isolated from photosynthetic sludge. Aquacultural Engineering, 19 (3), 1999, 179-193.

Junguo He, Guangming Zhang, Haifeng Lu, Treatment of soybean wastewater by a wild strain Rhodobacter sphaeroides and to produce protein under natural conditions, 4 (3), 2010, 334–339.

Kaewsuk J, Thorasampan W, Thanuttamavong M, Seo GT, Kinetic development and evaluation of membrane sequencing batch reactor (MSBR) with mixed cultures photosynthetic bacteria for dairy wastewater treatment. J Environ Manage, 91, 2010, 1161–1168.

Kanazawa A, Effects of docosahexaenoic acid and phospholipids on stress tolerance of fish, Aquaculture, 155, 1997, 131–137.

Kantachote D, Torpee S, Umsakul K, The potential use of anoxygenic phototrophic bacteria for treating latex rubber sheet wastewater, Electron J Biotechnol, 8, 2005, 314-323.

Kartikey Kumar Gupta, Kamal Rai Aneja and Deepanshu Rana, Current Status of Cow Dung as a Bioresource For Sustainable Development. Gupta, Bioresour. Bioprocess, 3, 2016, 28.

## Journal of Chemical and Pharmaceutical Sciences

Kim J.K and Lee B, Mass production of Rhodopseudomonas palustris as diet for aquaculture. Aquacul. Engin. 23, 2007, 281–293.

Kobayashi M and Kobayashi M, Waste remediation and treatment using anoxygenic photosynthetic bacteria. In Anoxygenic Photosynthetic Bacteria ed. Blankenship R.E, Madigan M.T and Bauer C.E. Dordrecht, The Netherlands, Kluwer Academic Publishers, 1995, 1269–1282.

Kobayashi M and Tchan Y.T, Treatment of industrial waste solutions and production of useful by-products using a photosynthetic bacterial method, Wat. Res, 7 (8), 1973, 1219-1224.

Koblizek M, Falkowski P.G, Kolber Z, Diversity and distribution of photosynthetic bacteria in the black sea. Deep Sea Res, 53 (17-19), 2006, 1934-1944.

Kosamu I.B.M, Obst M, The influence of picocyanobacterial photosynthesis on calcite precipitation, Int. J. Environ. Sci. Tech, 6 (4), 2009, 557-562.

Kuzmanova S, Dimitrovski A & Doneva D, Microbiology Skopj, 26, 1989, 159-164.

Loo PL, Vikineswary S, Chong VC, Nutritional value and production of three species of purple non-sulfur bacteria grown in palm oil mill effluent and their application in rotifer culture, Aquacult Nutr, 19, 2003, 895-907.

Madigan M.T, Martinko J.M and Parker J, Brock Biology of Microorganism, Prentice Hall, Upper Saddle River, 8, 2000, 891-921.

Madukasi E.I, Dai X.I, Chunhua H, Zhou J.J, Potentials of phototrophic bacteria in treating pharmaceutical wastewater, Int. J. Environ. Sci. Tech, 7 (1), 2010, 165-174.

Mahajan A, Dua S, Functional properties of repressed (Brassica campestris vartoria) protein isolates, J. Food Sci. Technol, 32, 1995, 162-165.

Maneewan Suwansaard, Production of Hydrogen and 5-Aminolevulinic Acid by Photosynthetic Bacteria from Palm Oil Mill Effluent Authors, Maneewan Suwansaard Description, Thesis (Ph.D, Biotechnology)--Prince of Songkla University, 2010.

Najafpour, Bhasem D, Single cell Protein, Biochemical Engineering and Biotechnology, 2007, 332-341.

Napavarn Noparatnaraporn and Shiro Nagai, Selection of Rhodobacter Sphaeroides P47 A Useful Source of Single Cell Protein, J. Gen. Appl. Microbiol, 32, 1986, 351-359.

Napavarn Noparatnaraporn, Savitr Trakulnaleumsai, Roberto Silveira G, Yoshinori Nishizawa, Shiro Nagai, SCP production by mixed culture of *Rhodocyclus gelatinosus* and *Rhodobacter sphaeroides* from Cassava Waste. Journal of Fermentation Technology, 65 (1), 1987, 11-16.

Nasseri AT, Rasoul-Amini S, Morowvat MH, Ghasemi Y, Single cell protein, Production and process, Ame. J. Food Technol, 6, 2011, 103-116.

Nastee Kornochalert, Duangporn Kantachote, Sumate Chaiprapat, Somkiet Techkarnjanaruk Use of *Rhodopseudomonas palustris* P1 stimulated growth by fermented pineapple extract to treat latex rubber sheet wastewater to obtain single cell protein, 64 (3), 2014, 1021-1032.

Neik T.S, The enhancement of growth and survival of ornamental koi (Cyprinus carpio) larvae using phototrophic bacteria, Master thesis, Faculty of Science, University of Malaya, Kuala Lumpur, Malaysia, 2006.

Nigam N.M, Cultivation of Candida langeronii in sugarcane bagasse hemi cellulose hydrolysate for the production of single cell protein, Microbiol W.J and biotechnol, 16, 2000, 367-372.

Noparatnaraporn N and Nagai S, Selection of *Rhodobacter sphaeroides* P47 as a useful source of single cell protein, J. Gen. Appl. Microbiol, 32, 1986, 351-359.

Noparatnaraporn N, Sasaki K, Nishizawa Y and Nagai S, Stimulation of vitamin B12 formation in aerobically-grown Rhodopseudomonas gelatinosa under microaerobic conditions, Biotechnol. Lett, 8 (7), 1986, 491-496.

Norbert Pfennig. *Rhodopseudomonas acidophila*, sp. n, a New Species of the Budding Purple Non sulfur Bacteria. Journal of Bacteriology, 99 (2), 1969, 597-602.

Okubo Y, Hiroyuki F, Akira H, Characterization of phototrophic purple non-sulfur Bacteria forming colored microbial mats in a swine wastewater Ditch. Appl. Environ. Microbiol, 72 (9), 2006, 6225-6233.

## Journal of Chemical and Pharmaceutical Sciences

Ponsano E.H.G, Lacava P.M and Pinto M.F, Isolation of Rhodocyclus gelatinosus from poultry slaughterhouse wastewater. Brazilian Archives of Biology and Technology, 45 (4), 2002, 1-8.

Poonsuk Prasertsan, Suteera Prasertsan and Aran H-Kittikun. Recycling of agro industrial wastes through cleaner technology, Biotechnology- Encyclopedia of life support system (EoLss), 10, 1997.

Prasertsan P, Choorit W and Suwano S, Isolation, identification and growth conditions of photysnthetic bacteria found in sea food processing waste water, World.J Microbio and Biotech, 9, 1993, 590-592.

Prasertsan P, Jaturapornpipat M and Siripatana C, Utilization and treatment of tuna condensate by photosynthetic bacteria. Pure and Applied Chemistry, 69 (11), 1997, 2439-2445.

Rainuzzo J.R, Reitan K.I, Olsen Y, The significance of lipids at early stages of marine fish, a review, Aquaculture 155, 1997, 103–115.

Ramchander Merugu M.S.K, Prasad S, Girisham and Reddy S.M, Tolerance of Certain Pesticides by two Nitrogen fixing Anoxygenic Phototrophic Bacteria Nat. Env. Pol. Tech, 7, 2008, 467-469.

Ramchander Merugu, MP Pratap Rudra, S Girisham, SM Reddy, Biotechnological applications of purple non sulphur phototrophic bacteria, A minireview, International Journal of Applied biology and Pharmaceutical technology, 3 (1), 2012, 376-384.

Saejung C, Thammaratana T, Biomass recovery during municipal wastewater treatment using photosynthetic bacteria and prospect of production of single cell protein for feedstuff, Environ Technol, 37 (23), 2016, 3055-3061.

Salma A.G, Miah K.M.A, Tareq, Maki and Tsuji H, Effect of dietary Rhodobacter capsulatus on egg-yolk cholesterol and laying hen performance, J. Nutrition, 2007.

Saquido P.M.A, Cayabyab V.A, Vyenco F.R, Bioconversion of banana waste into single cell protein, J. Applied Microbiol & Biotechnol, 5 (3), 1981, 321-326.

Sargent J, McEvoy L.A, Bell J.G, Requirements, presentation and sources of polyunsaturated fatty acids in marine fish larval feeds, Aquaculture, 155, 1997, 117–127.

Sasaki K, Noparatnaraporn N, Hayashi M, Nishizawa Y and Nagai S, Single cell protein production by treatment of soybean wastes with *Rhodopseudomonas gelatinosa*, J. Ferment. Technol, 59, 1981, 471-477.

Sasikala G.H And Ramana C.H.V, Biotechnological potentials of anoxygenic phototrophic bacteria. I. production of Single-Cell Protein, vitamins, ubiquinones, hormones, and enzymes and use in waste treatment, Advances in Applied Microbiology, 41, 1995, 173-226.

Sawada H, Parr RC, Roger PL, Photosynthetic bacteria in waste water treatment, J. Ferment. Technol, 55, 1977, 326-336.

Siefert E, Irgens R.L, Pfennig N, Phototrophic purple and green bacteria in a sewage treatment plant, Appl. Environ. Microb, 35 (1), 1978, 38-44.

Smith M.E, Bull A.T, Protein and other compositional analysis of Saccharomyces fragilis grown on coconut water waste, J. Applied Bacteriol, 41, 1976, 97-107.

Tan Kar Soon, Sujjat Al-Azad, and Julian Ransangan. Isolation and Characterization of Purple Non-Sulfur Bacteria, Afifella marina, Producing Large Amount of Carotenoids from Mangrove Microhabitats, J. Microbiol. Biotechnol, 24 (8), 2014, 1034–1043.

Tipparat H, Kittikun A.H, Optimization of single cell protein production from cassava starch using Schwanniomyces castellii. W.J. Microbiol. & Biotechnol, 11, 1995, 607-609.

Worowit Whagchenchom, Wilai Chien Chaisri, Pareena Tapaneeyawarorawong, Sorawit Powtongsook, Wastewater from Instant Noodle Factory as the Whole Nutrients Source for the Microalga Scenedesmus sp. Cultivation. Environ. Eng. Res, 19 (3), 2014, 283-287.

Yegani R, Satoshi Y, Kazunori M, Tomoshisa K, Shigeo K, Improvement of growth stability of photosynthetic bacterium Rhodobacter capsulatus, J. Biosci. Bioeng, 100 (6), 2005, 672-677.

Zhang X.X, Zhao D.Y, Wang Z. X, Wu B, Li W.X, Cheng S.P, Environmental biological model based on optimization of activated sludge process. Int. J. Environ. Sci. Tech, 6 (1), 2009, 69-76.

ISSN: 0974-2115

# www.jchps.com

# Journal of Chemical and Pharmaceutical Sciences

Zhao G, Zhang W, Zhang G, Production of single cell protein using waste capsicum powder produced during capsanthin extraction, Lett Appl Microbiol, 50, 2010, 187-191.

Zhu H, Ueda S, Asada Y and Miyake J, Hydrogen production as a novel process of wastewater treatment-studies on tofu wastewater with entrapped *R.sphaeroides* and mutagenesis. International Journal of Hydrogen Energy, 27 (11-12), 2002, 1349-1357.

Zubi W, Production of single cell protein from base hydrolyzed of date extract byproduct by the fungus Fusarium graminearum. M. Sc. Thesis, Benghazi, Garyounis University, 2005.