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ACUTE TOXICITY AND ETHOLOGICAL RESPONSES OF FRESHWATER FISH CHANNA PUNCTATA (BLOCH) EXPOSED TO HOUSEHOLD SYNTHETIC DETERGENT

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Abstract: The detergents are household chemical cleaning compounds used in a wide range of our daily life for diverse purposes. 'Ghari' is a common detergent used largely in India. The present study examined the effects of a commercial household synthetic detergent 'Ghari' on the mortality and behavioural changes in the freshwater fish *Channa punctata*. The median lethal concentration (LC_{50}) of this detergent was determined using probit analysis software (SPSS version 26) at 95% confidence limit. The 96h LC_{50} value for *Channa punctata* was found to be 20.121 mg/L. Toxicity increased significantly with higher detergent concentrations and longer exposure periods. Morphological changes observed after 96 hours of exposure included body and gill discoloration, damaged fins, and excessive mucus secretion. Behavioural alterations in detergent-exposed fish included erratic swimming, restlessness, aggression, hyperactivity, and frequent movement near the bottom of the tank. Further research is recommended to explore the physiological effects for a better understanding of environmental impacts.

Keywords: Acute toxicity, Channa punctata, Detergent, Ethological responses, Ghari, LC₅₀.

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INTRODUCTION

Detergents are complex mixtures of surfaceactive compounds or surfactants, builders and bleaches. The contaminations of aquatic ecosystem by detergents depend upon the amount and nature of their active ingredients. The active ingredients are also known as builders, which mostly consist of a mixture of sodium salts, borate, phosphates, carbonates, carboxymethyl cellulose, sulphates, silicate, perborate, etc. (Neil, 1985). Modern detergents possess high cleaning efficiency, primarily due to the presence of surfactants, which reduce the surface tension of water (Singh and Pandey, 2023).

The input of detergents particularly in freshwater aquatic ecosystem is causing pollution and the pollution is harmful for biodiversity (Singh *et al.*, 2023). This has reached a point of serious concern as reports suggesting their adverse effect on water quality, aquatic organisms including



fishes, eutrophication of natural waterbody due to presence of large amount of phosphates in detergents, reducing primary productivity of waterbody. It has been established that even insignificant amount of detergents reduce the resistance of fish enabling parasitic fungi to develop on them causing destruction of their fins. Detergents also impede the food source locating ability of fishes.

Some of the detergent constituents show high biological activity and has a tendency to accumulate in organisms, making adverse effects possible even at very low level of exposure (Prakash and Verma, 2018). Fishes are good bioindicator as these are very sensitive to changes in the aquatic medium and thus, play an important role in the monitoring of water pollution (Srivastava et al., 2010). The freshwater fish like Channa punctata (Bloch) is highly sensitive to the household detergents and other toxic chemicals (Goel et al., 2022). A number of researches have also been done regarding the impact of various toxic substances on fishes (Shukla and Arya, 2017; Prakash and Verma, 2020a). Besides, several workers like Prakash and Verma (2014 and 2020b), Masih (2021), Aborisade and Atuanya (2023) and so on worked a lot on the effect of different pesticides on fish biology.

Detergents together with their ingredients and metabolites constitute the biggest groups of anthropogenic pollutants. The ability to these compounds to form foam is a serious problem because the organic contaminants or pathogenic micro-organisms are accumulated in the foam thereby posing an epidemiological threat. In addition, foaming also reduces aeration and causes hypoxia (Vasanthi et al., 2013; Prakash and Singh, 2020; Tasmin et al., 2023).

In consideration of the above facts and keeping in view the lack of systematic and correlative details, the present study is meant to determine the LC₅₀ value of commercial detergent 'Ghari' and its effect on freshwater snake headed fish, Channa punctata, by using SPSS and also to study the ethological responses due to its toxic effects. The study will provide a baseline data that could be useful contribution in eco-toxicity risk assessment to the fish.

MATERIALS AND METHODS

Acclimatization of Test Fish

The freshwater fish, Channa punctata (Bloch) were collected from local freshwater bodies and the healthy fish of uniform length $(9.5\pm0.2 \text{ cm})$ and weight (10.0±5g) were selected for the acute toxicity test. Then these fishes were brought to the laboratory in plastic container to avoid injury during transportation. The collected fishes were washed with 1.0 % solution of KMnO₄ for 5 minutes to remove any dermal infection. Fish were acclimated to laboratory conditions for 10 days at room temperature (26 \pm 1.5°C) prior to experimentation. For the first week, the acclimatized fishes were given artificial air by aerator. All of the fish were given commercial food pellets on a regular basis, and the medium (tap water) was changed every two days to get rid of food remnants and faeces. Feeding was stopped 24 hour prior to the toxicity test.

Determination of Acute toxicity (LC₅₀)

The experiments were carried out in glass aquaria of 15 litre capacity in laboratory conditions. The stock solution was prepared by dissolving 'Ghari' detergent in tap water. The range finding tests or exploratory tests were conducted to ascertain the final concentrations to be used in definitive test by following the standard method (APHA, 2005). Prior to the 96h static bioassay test, a rangefinding test was carried out to determine the range of concentrations to be used. The range finding test was conducted using a broad concentration range (1, 10, 20,and 100 mg/L) and the test was terminated after 24 h. Concentrations of the detergent that caused death within 30 minutes were not considered for the further definitive toxicity test/ LC_{50} test.

In the definitive test, the concentrations selected were based on the mortality values obtained with the range-finding test and were in appropriate logarithmic dilution series. Three replicate test tanks for each concentration (5, 10, 15, 20, 25, 30, 35 and 40 mg/L) contained a total of 5.0 L of the test solution and 10 test fishes. The percentage of mortality of the test fish after 96 hours exposure to each concentration was calculated separately using the formula: mortality (%) = no dead/ total number tested×100%. The dead fishes were removed immediately from the aquaria. The control experiment contained the tap water. LC_{50} value of detergent for freshwater fish, *Channa punctata* was determined by using probit analysis software (SPSS version 26) at 95% confidence limit.

RESULTS AND DISCUSSION

Fish acute toxicity studies play a vital role in environmental risk assessment and hazard classification, as they provide preliminary estimates of the relative toxicity of different chemicals across various species. In the present study, acute toxicity tests usually provide estimates of the exposure concentration of 'Ghari' detergent causing 50 % mortality (LC_{50}) to test species *Channa punctata* within 96 hours of exposure.

It is clear from the data that detergent toxicity was increased with increasing the concentrations of

detergents. At 96 h, when the test was terminated, mean mortality in response to 12 mg/L, 14 mg/L, 16 mg/L, 18 mg/L, 20 mg/L, 22 mg/L, 24 mg/L, 26 mg/L and 28 mg/L of 'Ghari' detergent was 10%, 20%, 20%, 30%, 40%, 50%, 70%, 80% and 100%, respectively (Table 1). The LC₅₀ value of the 'Ghari' detergent to Channa punctata was found to be 26.215 mg/L, 24.479 mg/L, 21.927 mg/L and 20.121 mg/L for an exposure period of 24, 48, 72 and 96 hr, respectively (Table 2). The 96h LC₅₀ value of Surf excel and Nirma for fresh water fish Mystus montanus were 20.0 mg/litre and 23.5 mg/litre respectively (Chandanshive, 2014). The LC₅₀ value of the common household synthetic detergent to Catla catla was found to be 14.20 ppm for an exposure period of 96 h (Arivizhivendhan et al., 2014). The Fig 1a-1d shows that mortality percentage was increased significantly with increase in log concentration of Ghari detergent and exposure period. The same kind of toxicity effect was observed by some workers (Bhaskaran, 1991; Maruthanayagam et al., 1994).

Table 1: Percentage of mortality of Fish, *Channa punctata* (Bloch) at different concentrations of 'Ghari' detergent (mg/L).

Concentration of detergent (ml/L)	Log conc. of detergent	No. of Test fish	Percent mortality at different exposure period (hours)			
			24	48	72	96
Control (0.0)	-	10	0	0	0	0
10	1.000	10	0	0	0	0
12	1.079	10	0	0	0	10
14	1.146	10	0	0	10	20
16	1.204	10	0	10	10	20
18	1.255	10	10	20	20	30
20	1.301	10	10	20	30	40
22	1.342	10	20	30	40	50
24	1.380	10	30	40	60	70
26	1.415	10	40	50	70	80
28	1.447	10	50	60	90	100
30	1.477	10	70	80	100	-
32	1.505	10	90	100	-	-
34	1.531	10	100	-	-	-

The alterations in the behavioural pattern are the most sensitive indication of potential toxic effects. The behavioural alteration observed in fish, Channa punctata increased with the dose and duration. The fish exposed to 10.0 to 34.0 mg/L detergent concentrations exhibited

Table 2: LC₅₀ values with 95% confidence limits for Mystus vittatus (Bloch) exposed to 'Ghari' detergent at different exposure period.

Exposure period (hrs.)	LC ₅₀ values of detergent (mg/L)	95% Confidence limits (v/v%)		Regression	Chi Square	Coefficient
		Lower limit	Upper limit	equation	value (P value)	of deter- mination (R ² Linear)
24	26.215	24.610	28.045	Y=-13.89+9.78X	4.151 (0.965)	0.971
48	24.479	22.686	26.684	Y=-9.79+6.99X	4.234 (0.936)	0.941
72	21.927	20.325	23.721	Y=-11.17+8.32X	3.862 (0.920)	0.926
96	20.121	18.329	22.269	Y=-7.89+9.35X	4.085 (0.849)	0.935

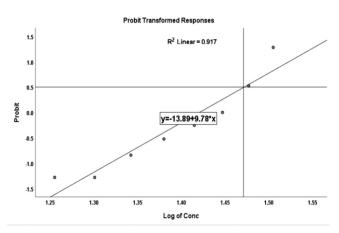


Fig. 1a: Regression line (based on Probit analysis) of Log concentration of 'Ghari' detergent vs % mortality of Channa punctata (at 24 hrs).

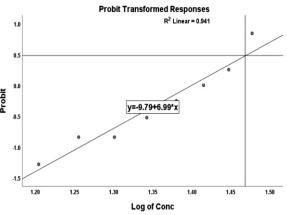


Fig. 1b: Regression line (Based on Probit analysis) of Log concentration of 'Ghari' detergent vs % mortality of Channa punctata (at 48 hrs).

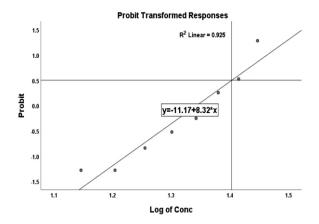


Fig. 1c: Regression line (Based on Probit analysis) of Log concentration of 'Ghari' detergent vs % mortality of Channa punctata (at 72hrs).

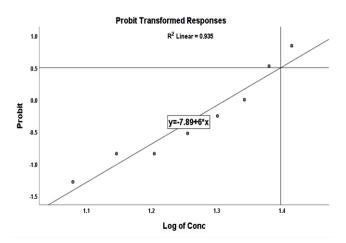


Fig. 1d: Regression line (Based on Probit analysis) of Log concentration of 'Ghari' detergent vs % mortality of Channa punctata (at 96 hrs).

abnormal behaviour in the form of erratic swimming, loss of equilibrium and enhanced surfacing behaviour. At the start of the exposure, the fish exposed to the detergent became alert; with the progression of the experiment, the fish stopped swimming and remained in static position in response to the sudden changes in the surrounding environment. Authors observed that the fish experienced progressive surfacing, air gulping, restlessness, escaping movement, erratic swimming, increased mucous secretion lethargic, loss of equilibrium, difficulty in respiration, exhibited convulsions, settling to the bottom before death. There were no behavioural changes observed in the control group.

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REFERENCES

- 1. Aborisade W.T. and Atuanya E.I. (2023). Ecotoxicological studies of pesticide formulations on soil sentinel's microflora (*Nitrosomonas* and *Nitrobacter* spp.) and mesofauna (*Ephyriodrilus* spp.). *International Journal of Biological Innovations*. 5(1): 01-13. https://doi.org/10.46505/IJBI.2023.5101
- **2. APHA** (2005). Standard methods for the examination of water and wastewater. 21th edn., Washington D.C.
- 3. Arivizhivendhan K.V., Ali J., Chitrarasu P., Sampath R.K., Boopathy R., Mahesh M. and Regina R.M. (2014). The Sub-lethal effect of detergent on the biochemical parameter of Fresh water fish, Catla catla. International Journal of Pharmaceutical & Biological Archives. 5(2): 145-149.
- 4. Bhaskaran P., Gopalakrishnaswamy G. and Sathyabama (1991). Impact of commercial detergent (Nirma) on feeding energetic and protein metabolism in the teleost *Orechromis mossambicus*. *J. Ecotoxical*. *Environ*. *Monit*. 1 (1): 31-40.
- 5. Chandanshive N.E (2014). Effects of different concentrations of detergents on Dissolved Oxygen consumption in fresh water fish *Mystus montanus*. *Int. Res. J. Environment Sci.* 3(8): 1-5.

- 6. Goel R., Arya S. and Singh R. (2022). Impact of pyrethroid insecticides on the haematology of *Channa punctata* (Bloch). *International Journal on Biological Sciences*. 13(2): 109-111. https://doi.org/10.53390/ijbs.v13i2.2
- 7. Maruthanayagam C., Arun S. and Suramaniyan P. (1994). Acute toxicity of synthetic detergent to *Macrobrachiumla* merrei. Adv. in . Zool. Series. 1: 165-170.
- 8. Masih S.C. (2021). Impact of Monocrotophos pesticide on serum biochemical profile in freshwater fish, *Cirrhinus mrigala* (Hamilton, 1822). *International Journal of Biological Innovations*. 3(2):402-406. https://doi.org/10.46505/IJBI.2021.3222
- **9. Neil O.P.** (1985). Environmental Chemistry. Allen and Unwin, London, U.K.
- **10. Prakash S. and Singh D.** (2020). Impact of distillery effluent on Behaviour and Oxygen consumption of *Cyprinus carpio* (L.). *International journal of Scientific Research in Biological Sciences*. **7**(3): 34-37.
- 11. Prakash S. and Verma A.K. (2014). Effect of Organophosphorus Pesticide (Chlorpyrifos) on the Haematology of *Heteropneustes fossilis* (Bloch). *International Journal of Fauna and Biological Studies*. 1(5): 95-98.
- **12. Prakash S. and Verma A.K.** (2018). Effect of synthetic detergent on biochemical constitutions of freshwater major carp, *Labeo rohita*. *International Journal on Agricultural Sciences*. 9(1): 57-60.
- 13. Prakash S. and Verma A.K. (2020a). Effect of Arsenic on Serum Biochemical parameters of a fresh water cat fish, *Mystus vittatus*. *International Journal of Biological Innovations*. 2 (1): 11-19. https://doi.org/10.46505/IJBI.2020.2102
- 14. Prakash S. and Verma A.K. (2020b). Effect of organophosphorus pesticides on Biomolecules of fresh water fish, Heteropneustes fossilis (Bloch). Indian Journal of Biology. 7(2): 65-69. http://dx.doi.org/10.21088/ijb.2394.1391.7220.8
- **15. Shukla M. and Arya S.** (2017). Cadmium toxicity induced morphological alteration in

- indigenous fish Heteropneustes fossilis (Bloch.). Green Chemistry and Technology Letters. 3(1):21-25. 10.18510/gctl.2017.315
- 16. Singh P. and Pandey R.K. (2023). Alteration in biochemical and haematological parameters on exposure to household detergent (tide) of the Asian snakehead fish Channa punctata. International Journal of Biological Innovations. 5(2): 01-11. doi.org/10.46505/ IJBI.2023.5201
- 17. Singh R., Verma A.K. and Prakash S. (2023). The web of life: Role of pollution in biodiversity decline. International Journal of Fauna and Biological Studies. 10(3): 49-52. 10.22271/23940522.2023.v10.i3a.1003
- 18. Srivastava A.K., Mishra D., Shrivastava S., Srivastav S.K. and Srivastav A.K. (2010). Acute toxicity and behavioural responses of

- Heteropneustes fossilis to an organophosphate insecticide, Dimethoate. International Journal of Pharma and Bio Sciences. 1(4): 359-363.
- 19. Tasmin R., Rifa M. and Zari S.N. (2023). Acute toxicity of two detergents on the juvenile climbing perch (*Anabas testudineus*) and stinging catfish (Heteropneustes fossilis). Bangladesh J. Zool. 51(1): 87-98. https://doi.org/10.3329/bjz.v51i1.68659
- 20. Vasanthi J., Binukumari S. and Saradhamani N. (2013). A study on the effect of detergent suryagold on the chronic toxicity of the freshwater fish, Cirrhinus mrigala. IOSR Journal of Environmental Sciences, Toxicology and Food Technology. 7(6):18-20.