

Content available at: https://www.ipinnovative.com/open-access-journals

IP Archives of Cytology and Histopathology Research

Journal homepage: https://www.achr.co.in/



Original Research Article

Observational study on serum markers and circulating tumor cells in ovarian cancer

Pravesh Gupta¹, Riddhi Jaiswal^{1,*}, Piyush Tripathi², Vimala Venkatesh²

¹Dept. of pathology, King George's Medical University, Lucknow, Uttar Pradesh, India



ARTICLE INFO

Article history: Received 26-07-2021 Accepted 21-08-2021 Available online 16-09-2021

Keywords:
Ovarian lesion
Circulating tumor cells
Serum CA125
Serum HE4

ABSTRACT

Aims: 1) To detect Cyclophilin, APC and SFRP5 genes associated with Epithelial ovarian cancer by PCR. 2) To study and compare the prognostic and diagnostic efficacy of serological markers like Ca 125 and HE4 and their correlation with epithelial/ non epithelial ovarian neoplasms.

Settings and Design: Comparative observational study, Prospective study

Methods and Materials: 64 cases fulfilling the inclusion criteria and giving their consent for inclusion in the study were enrolled as subjects of the present study over a period of one year. After DNA extraction (Invitrogen mini kit, USA) conventional PCR to amplify the extracted DNA and further subjected them to agarose gel electrophoresis for the identification(expression) of 3 genes i.e Cyclophillin, APC and SFRP5, was done; However, none expressed.

ELISA was used to assess CA125 and HE4 pre and post surgical intervention.

Results: The serum markers were raised more in malignant epithelial ovarian cancer cases and levels plummeted after surgical intervention, as compared to benign masses.

We could not establish correlation of the genes' expression with the serum markers and histopathology.

Conclusions: Combining HE4 and CA125 both might be more helpful than either of them using alone, in diagnosing as well as prognosticating ovarian diseases.

A panel of multiple genes on a larger sample size may be needed for CTC detection.

Key Messages: Early detection of ovarian tumors leads to early diagnosis and hence early institution of intervention and hence decreased morbidity and mortality.

This is an Open Access (OA) journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: reprint@ipinnovative.com

1. Introduction

Ovarian malignancies constitute 3% of all malignancies in the females and is the fifth most common cause of death due to cancer in women. ¹ In most of the population-based cancer registries in India, ovarian cancer is the third leading site of cancer among women, trailing behind cervix and breast cancer. The age-adjusted incidence rates of ovarian cancer vary between 5.4 and 8.0 per 100,000 population in different parts of India. ²

E-mail address: riddhiadvay@gmail.com (R. Jaiswal).

Primary ovarian tumors may arise from surface epithelial cells, germ cells, or sex cord stromal cells, so have been classified accordingly. Secondary tumours are implanted on to the ovary from somewhere else.

Dualistic model of epithelial ovarian cancer (EOC), which divides epithelial ovarian carcinomas into two broad categories, designated type I and type II. Prototypic type I tumour is low grade serous serous carcinoma, which has a high frequency of KRAS and BRAF mutations but no TP53 mutations and prototypic type II tumor is high grade serous carcinoma which is characterized by high level of genetic instability and harbours TP53 mutations in nearly all cases.

²Dept. of Microbiolofy, King George's Medical University, Lucknow, Uttar Pradesh, India

^{*} Corresponding author.

The main problem with the epithelial ovarian cancers is that these patients (almost 75%) are diagnosed on advanced stages (FIGO III/IV). By the time these are diagnosed, metastases have already occurred by transperitoneal, hematogenous and lymphatic routes.^{3,4}

1.1. Diagnostic tools

1.1.1. Molecular methods

A. Polymerase Chain Reaction: Molecular diversity exists within specific histological subtypes of EOC, between different tumors of an individual patient, as well as within individual tumors. EOC are classified as either type I or type II tumor on molecular basis with implications on disease pathogenesis and prognosis. Association of various genes alteration with the EOC I have been studied like BRAF, KRAS PIK3CA, CTNNB1, & CDKN2A and type II ovarian malignancies with BRCA1, BRCA2, MYC, MECOM, CCNE1, PRCK1, NOTCH3, KRAS, ERBB2, KIT & EGFR. ^{5–8}

For the early diagnosis of EOC, multiplex nested methylated specific polymerase chain reaction (PCR) is another method to detect the methylation level of serum and tissue samples in patients with ovarian carcinoma and the results compared with the detection with CA125. 9

B. Circulating Tumor Cells (CTC) and Cell free DNA (cfDNA): Tumor-derived DNA has been identified in the body fluids of patients with a variety of cancers, including colorectal, head and neck, lung, bladder, kidney, and prostate. ^{10–14} Identifying tumor-specific molecular alterations in urine, saliva, sputum, and stool can be a noninvasive diagnostic test for cancer. ^{15–18}

Liquid biopsy, involves the collection and analysis of circulating tumor cells (CTCs), circulating tumor DNA (ctDNA), circulating cell-free microRNAs (cfmiRNAs), and exosomes. ^{16,17}

The use of cfDNA as a screening method has been evaluated in cancers of renal cell, lung, testicular, colorectal, and breast origin, suggesting that it could be used as a non-invasive biomarker for a wide variety of cancer types. ^{18–21}

These cfDNAs were found to range between 0.5 to 21 kb in length, with a plasma concentration ranging from 0.15 to 12 mg/mL, used as a diagnostic tool to detect different forms of cancer including hematological, colorectal and thoracic neoplasms. This was followed by studies that focused on identifying micrometastases in form of CTCs. 23–25 Bettegowda et al. showed that these micrometastases signal can also be identified by measuring ctDNAs. 25

1.1.2. Serum markers

A. CA125 (Carbohydrate or Carcinoma Antigen) is a high molecular weight glycoprotein. Serum CA125 levels are routinely ordered in women with adnexal masses, and levels less than 35 U/ml are considered to be normal. However, CA125 has low specificity as it is also increased in benign

endometriosis, endometrial and cervical adenocarcinoma, breast and colon cancer etc. ^{26,27}

B. Serum HE4 (Human Epididymis protein) is used to monitor recurrence/progressive disease in patients with epithelial ovarian malignancy. Normally female genital tract and epididymal epithelium secrete HE4. Its over expression is seen in serous and endometrioid ovarian malignancies. ²⁶

1.2. Steps and methods

1.2.1. Study design
Comparative observational study.

1.2.2. Sample size 64 (95% CI; 5% error; prevalence of 5%).

1.2.3. Duration of study

One year.

The study was carried out in the department of Pathology, in collaboration with Microbiology and Obstetrics and Gynaecology, was approved by the Institutional Ethical Committee and review board. Patients of clinically diagnosed and radiologically suggested ovarian mass were included in study. Final histopathological diagnosis was recorded in all cases.

Patients with a history of cancer, either gynecological or from other systems or inflammatory diseases as a side diagnosis were excluded.

The blood samples were collected twice, once before any invasive procedures or before start of any medical treatments, and second between 4^{th} - 7^{th} day post surgery. Clinical information including demographic information, serum CA 125 and HE4 levels, tumor stage, histology, and ultrasonographic finding was obtained from the patients' medical records and pathology reports. Serum markers were tested and analysed in pre-operative and post-operative samples, in all cases.

The tumor volume in epithelial ovarian cancers was obtained before any tumor reduction by Radiology. CA 125 was measured before any treatment or surgery. The cut off value of 35 U/mL was used to distinguish between normal and pathologic cases.

1.3. Sample collection, storage and steps

A total of 5 mL of venous blood sample were collected. These samples were transferred to an 8 mL gel serum separator tube (BD—Becton, Dickinson and Company) and kept at room temperature for coagulation. Subsequently, the samples were centrifuged for 10 min at 1000 g. The samples were processed within 2 hours after collection to prevent loss of cell-free DNA for PCR. ^{23,24}

The serum of all patients were collected and DNA was extracted by Invitrogen DNA Mini Kits (USA, Invitrogen company), with all operations being conducted strictly

according to the kit instructions. PCR was performed for 3 genes that is Cyclophilin, SFRP5 and APC.

Steps performed were DNA Extraction, Preparation of lysate: (Manufacturer's instructions were followed), Binding of DNA, Washing of DNA, Elution of DNA and finally Agarose gel electrophoresis was performed to analyse results.

1.3.1. Conventional PCR cycles and thermal conditions Conditions used for PCR were "initial denaturation at 95°C for 10 min followed by 40 cycles of 1min denaturation at 95°C, 1min annealing at 55°C, and 1min elongation at 72°C. The last cycle was followed by a final extension of 10min at 72°C.

PCR was performed with prepublished primers for 3 genes, cyclophilin with forward sequence 5'-TTCTTCATCACCTATGGCAAAC-3' & reverse primer 5'-GCAACTTCTCCAACTCATCTAG-3', SFRP5 WITH **FORWARD** primer 5'-CAGATGTGCTCCAGTGACTTTG-3' & reverse primer 5'-AGAAGAAAGGGTAGTAGAGGGAGand APC with forward primer 5'-GAGACAGAATGGAGGTGCTGC-3' &reverse primer 5'-GTAAGATGATTGGAATTATCTTCT-3'. PCR products were identified by 2% agarose gel electrophoresis (AE), and DNA was regarded to be qualified when there were significantly visible products. (seephotoplates).

1.3.1.1. Agarose gel electrophoresis. To visualize, 5μ l of the PCR amplicon was loaded with gel loading dye (Thermo Fisher Scientific, India) in 1.5 % agarose gel containing 0.5μ l/ml of ethidium bromide (0.5mg/ml, Medox biotech India Pvt Ltd) along with molecular weight marker (100bp DNA ladder; Bangalore Genei, India) followed by electrophoresis at 80 V for 2 h and multiple amplified DNA was analysed by 264nm wavelength UV transilluminator and gel was documented.(images attached at photoplates).

A sample would be labelled as CTC positive if at least one of 3 gene marker panel is found to be over expressed.

1.3.2. Detection of serum CA125

Serum CA125 concentration was detected by ELISA CA125 Test principle: The CA125 quantitative test is based on a solid phase enzyme linked immunosorbent assay, measured spectrophotometrically at 450nm. The concentration of CA125 is directly proportional to intensity of the sample.

1.3.3. Detection of Human Epididymal Protein 4 Serum HE4 was detected by sandwich ELISA

1.3.3.1. Testprinciple. The optical density (OD) measured with spectrophotometry at a wavelength of 450 nm \pm 2 nm is proportional to the concentration of Human HE4 calculated in samples by comparing the OD of the samples with the

standard curve.

2. Results

64 cases fulfilling the inclusion criteria and giving their consent for inclusion in the study were enrolled as subjects of the present study. However, 5 samples were discarded due to either inadequacy or hemolysis. All the calculations are based on 59 samples studied for all parameters.

On the Serum samples which we procured, after DNA extraction (Invitrogen mini kit, USA) we performed conventional PCR to amplify the extracted DNA and further subjected them to electrophoresis for the identification(expression) of 3 genes i.e Cyclophillin, APC and SFRP5, however, none of the genes expressed in either benign or malignant cases, in either pre or post operative samples.

Age of patients enrolled in the study ranged between 12 to 85 years, mean age of patients was 38.24 ± 16.06 years. Majority of the cases enrolled in the study were aged above 30 years (66.1%).

50(84.7%) were married and only 9(15.3%) cases were unmarried.

Majority of the cases enrolled in the study were multipara to grandmultipara (P2 and above; n=39, 66.1%), only 6.8% were Unipara and rest were Nullipara (27.1%).

Size of lesions of the patients enrolled in the study ranged from 1.78 to 31.0 cm, mean size of lesions was found to be 8.64 ± 5.91 cm. Only 35.6% cases had lesion size \leq 5 cm, 32.2% each of the patients had lesion size 5-10 cm and >10 cm.

Majority of the patients had unilateral involvement (71.2%) rest 28.8% had bilateral involvement.

Cystic echotexture on imaging was observed in majority of the cases (54.2%), solid echotexture was observed for 13.6% in rest 32.2% cases mixed echotexture was observed.

Pre-operatively, patients with Malignant lesions as compared to Benign lesions had significantly higher CA-125 levels (902.39±1792.10 vs. 62.49±109.91 U/ml) as well as higher HE-4 levels (744.31±405.21 vs. 72.68±69.69 pmol/l). Table 2A.

Post-operatively too, patients with Malignant lesions as compared to Benign lesions had significantly higher CA-125 levels (54.09±39.93 vs. 27.60±25.86 U/ml) as well as higher HE-4 levels (150.11±88.20 vs. 35.82±28.02 pmol/l). Table 2B.

Decline in pre-operative CA-125 and HE-4 was observed in patients with benign as well as malignant lesions. Table 2C.

Changes were found to be significant statistically for both the markers, in benign lesions while significant statistically only for HE-4 marker, in malignant lesionsFigure 1

On receiver-operator characteristic analysis, the area under curve was found to be 0.782 for CA-125 and 0.926 for HE-4. The difference between two was not significant

Table 1: Histopathological Diagnosis

S.No	Diagnosis		No.	%
	Benign		43	72.9
	Cystic lesion	5/43 (11.6%)	5	8.5
	Serous cyst	15/43 (34.9%)	15	25.4
1.	Mature cystic	7/43 (16.3%)	7	11.9
	Follicular cystic	5/43 (11.6%)	5	8.5
	Hemorrhagic cyst	4/43 (9.34%)	4	6.8
	Others (Benign)	7/43 (16.3%)	7	11.9
	Malignant		16	27.1
2.	Serous cystadenoCa	10/16 (62.5%)	10	16.9
	Others (Malignant)	6/16 (37.5%)	6	10.2

Table 2: A. Evaluation of mean preoperative HE-4 and CA-125 levels for their diagnostic efficacy

Diamentia dameia	N T	CA-12:	5 (U/ml)	HE-4 (pmol/l)		
Diagnostic domain	N	Mean	SD	Mean	SD	
Benign vs Malignant						
Benign	43	62.49	109.91	72.68	69.69	
Malignant	16	902.39	1792.10	744.31	405.21	
Statistical significance		't'=3.104; p= 0.003 (Sig)		't'=10.603; p<0.001		

Dia autia da ai		CA-125	5 (U/ml)	HE-4 (pmol/l)		
Diagnostic domain	n	Mean	SD	Mean	SD	
Benign vs Malignant						
Benign	43	27.60	25.86	35.82	28.02	
Malignant	16	54.09	39.93	150.11	88.20	
Statistical significance		't'=2.994; p=0.004 (Sig)		't'=7.616; p<0.001		

Table 2C: Change in Pre-Operative HE4 and CA-125 levels (Paired 't' test)										
		C	A-125 (U/m	1)				HE-4 (pmol/	(I)	
	Mean Ch.	SD	% Ch.	't'	ʻp'	Mean Ch.	SD	% Ch.	't'	'p'
Benign	-34.89	91.56	-55.83	-2.499	0.016	-36.85	53.43	-50.70	-4.523	< 0.001
Malignant	-848.3	1766.3	-94.01	-1.921	0.074	-594.2	347.1	-79.83	-6.848	< 0.001

Table 2D Comparative parameters of HE4 and CA125 at the expected cut off values (Receiver Operator Characteristic Curve)							
Parameter	Area under curve (95% CI)	Projected cut-off value	Projected Sensitivity	Projected Specificity			
CA-125	0.782 (0.63-0.94)	≥92.35	75.0%	81.4%			
HE-4	0.926 (0.84-1.01)	≥102.62	87.5%	70.1%			

statistically (p>0.05). Table 2D.

3. Discussion

3.1. Molecular methods

On evaluating the regressed coordinates for different sensitivity/specificity trade-off, the cut-off value >92.35 U/ml for CA-125 and or HE-4 and cut-off value >102.62 p/mol. Sensitivity and specificity for CA-125 were found to be 75.0% & 81.4% while that for HE-4 were 87.5% & 70.1% in differentiating malignant masses and benign lesions.

Since none of our samples came out to be positive for either of the three genes (cyclophyllin, SFRP5 and APC), we probed further into literature to compare our results. Most of the related studies conclude that concentration of CTCs presented in ovarian cancer are extremely low, $1/10^9$ blood cells or $1/10^6$ nucleated blood cells, and hence difficult to detect. Moreover stringent procurement, storage and test conditions are demanded.³

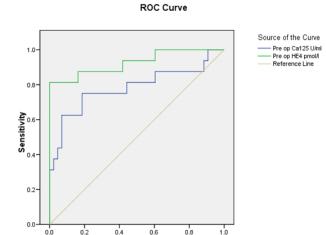


Fig. 1:

1 - Specificity

Board et al in 2010, to detect SNV (PIK3CA) alteration by allele specific PCR in operable stage of breast cancer reported 0% (0/14) sensitivity in patients with detected ctDNA with marker positive breast cancers.²⁸

However Shao et al. in 2015 suggested that there is a definite correlation between levels of serum cfDNA and development of ovarian cancer, but their detection is challenging.²⁹

Ignatiadis et al. in 2015 said that ctDNA may represent an exceedingly rare subpopulation within total cell-free DNA, at levels corresponding to one genome equivalent in 5 mL of plasma (0.01% allele fraction), and may be undetectable in plasma volumes typically sampled. 30

3.2. Serum markers

CA-125 levels ranged from 6.10 to 7087 units. A mean of 62.49 units/ml and SD of 109.91 in benign cases, and a mean of 902.39 units/ml and SD of 1792.10 in malignant cases was calculated, as is shown in Table 2A. The ability of CA125 in differentiating benign from malignant cases was found to be statistically significant (p=0.003). Along with USG findings, it proves to be a sensitive method to differentiate benign from malignant masses. Some studies however show that CA125 can sometimes over-diagnose benign cases like endometriosis and hence lead to unnecessary operation.

Human Epididymis 4 levels: HE4 levels ranged from 10.04 pmole/litre to 1391.08 pmole/litre.

In benign cases, a mean of 72.68 and a standard deviation of 69.69 was obtained while in malignant cases, mean was 744.31 and SD was 405.21. HE-4 lvels were found to be significantly higher in malignant cases as compared to benign (p<0.001).

4. Conclusion

Detection of circulating tumor DNA in ovarian cancer sera is difficult, and requires meticulous draft of multiple genes in panel to get a significant yield Serum markers must be used in conjunction with clinico-radiological findings.

5. Acknowledgement

I would like to express the deepest gratitude to my chief supervisor and guide Dr. Riddhi Jaiswal (M.D.) Associate Professor, Department of Pathology, KGMU, Lucknow for her constant encouragement, support and constructive criticism throughout the course of this study. I have been fortunate as I got opportunity to work under her supervision. Throughout the thesis, she handled my careless mistakes with great patience and bestowed her motherly love on me. I thank you mam for everything. Dr. Riddhi Jaiswal has been my mentor, my guide, my icon. In true sense words fail to express my heartfelt thanks.

I am highly grateful to Family and friends who supported me throughout and in most difficult times.

I am very much thankful to the patients for their cooperation and wish them good health.

6. Conflict of Interest

The authors declare that there are no conflicts of interest in this paper.

7. Source of Funding

None.

References

- Zhang Q, Hu G, Yang Q, Dong R, Xie X, Ma D, et al. A multiplex methylation-specific PCR assay for the detection of earlystage ovarian cancer using cell-free serum DNA. *Gynecol Oncol*. 2013;130(1):132–9. doi:10.1016/j.ygyno.2013.04.048.
- Basu P, De P, Mandal S, Ray K. Biswas J2 Study of 'patterns of care' of ovarian cancer patients in a specialized cancer institute in Kolkata, eastern India. *Indian J Cancer*. 2009;46(1):28–33. doi:10.4103/0019-509x.48592.
- Cheng X, Zhang L. Yajuan Chen and Chen Qing Circulating cellfree DNA and circulating tumor cells, the "liquid biopsies" in ovarian cancer. *J Ovarian Res*. 2017;10. doi:10.1186/s13048-017-0369-5.
- Zhang X, Lia H, Yua X, Li S, Lei Z, Li C, et al. Analysis of Circulating Tumor Cells in Ovarian Cancer and Their Clinical Value as a Biomarker. *Cell Physiol Biochem*. 2018;48(4):1983–94. doi:10.1159/000492521.
- Rojas V, Hirshfield KM, Ganesan S. Molecular Characterization of Epithelial Ovarian Cancer: Implications for Diagnosis and Treatment. *Int J Mol Sci.* 2016;17(12):2113. doi:10.3390/ijms17122113.
- Singer G, Oldt R, Wang BG, Sidransky D, Kurman RJ. Shih Ie M. Mutations in BRAF and KRAS characterize the development of low-grade ovarian serous carcinoma. *J Natl Cancer Inst.* 2003;95(6):484–6. doi:10.1093/jnci/95.6.484.
- Jones S, Wang TL, Kurman RJ, Nakayama K, Velculescu VE, Vogelstein B, et al. Low-grade serous carcinomas of the ovary contain very few point mutations. *J Pathol*. 2012;226(3):413–20. doi:10.1002/path.3967.

- 8. Hunter SM, Anglesio MS, Ryland GL, Sharma R, Chiew YE, Rowley SM, et al. Molecular profiling of low grade serous ovarian tumours identifies novel candidate driver genes. *Oncotarget*. 2015;6:37663–77. doi:10.18632/oncotarget.5438.
- Umetani N, Kim J, Hiramatsu S. Increased integrity of free circulating DNA in sera of patients with colorectal or periampullary cancer: direct quantitative PCR for ALU repeats. *Clin Chem.* 2006;52(6):1062–9. doi:10.1373/clinchem.2006.068577.
- Sidransky D, Tokino T, Hamilton SR, Kinzler KW, Levin B, Frost P, et al. Identification of ras oncogene mutations in the stool of patients with curable colorectal tumors. *Science*. 1992;256(5053):102–5. doi:10.1126/science.1566048.
- Fliss MS, Usadel H, Caballero OL, Wu L, Buta MR, Eleff SM, et al. Facile detection of mitochondrial DNA mutations in tumors and bodily fluids. *Science*. 2000;287(5460):2017–9.
- 12. Sidransky D, Eschenbach AV, Tsai YC, Jones P, Summerhayes I, Marshall F, et al. Identification of p53 gene mutations in bladder cancers and urine samples. *Science*. 1991;252(5006):706–9. doi:10.1126/science.2024123.
- Cairns P, Esteller M, Herman JG, Schoenberg M, Jeronimo C, Sanchez-Cespedes M, et al. Molecular detection of prostate cancer in urine by GSTP1 hypermethylation. *Clin Cancer Res*. 2001;7(9):2727– 30.
- 14. Swisher EM, Wollan M, Mahtani SM, Garcia R, Goff BA, King MC, et al. Tumor-specific p53 sequences in blood and peritoneal fluid of women with epithelial ovarian cancer. *Am J Obstet Gynecol*. 2005;193(3):662–7. doi:10.1016/j.ajog.2005.01.054.
- Mellby LD, Nyberg AP, Johansen JS, Wingren C, Nordestgaard BG, Bojesen SE, et al. Serum Biomarker Signature-Based Liquid Biopsy for Diagnosis of Early-Stage Pancreatic Cancer. *J Clin Oncol*. 2018;36(28):2887–94. doi:10.1200/JCO.2017.77.6658.
- Giannopoulou L, Zavridou M, Kasimir-Bauer S, Lianidou ES. Liquid biopsy in ovarian cancer: the potential of circulating miRNAs and exosomes. *Transl Res.* 2019;205:77–91. doi:10.1016/j.trsl.2018.10.003.
- 17. Gang F, Guorong L, An Z, Gentil-Perret A, Christian G, Jacques T, et al. Prediction of Clear Cell Renal Cell Carcinoma by Integrity of Cell-free. *Urology*. 2010;75(2):262–5. doi:10.1016/j.urology.2009.06.048.
- Deligezer U, Eralp Y, Akisik EE, Akisik EZ, Saip P, Topuz E, et al. Size distribution of circulating cell-free DNA in sera of breast cancer patients in the course of adjuvant chemotherapy. Clin Chem Lab Med. 2008;46(3):311–7. doi:10.1515/CCLM.2008.080.
- Schwarzenbach H, Stoehlmacher J, Pantel K, Goekkurt E. Detection and monitoring of cell-free DNA in blood of patients with colorectal cancer. *Ann N Y Acad Sci.* 2008;1137:190–6. doi:10.1196/annals.1448.025.
- No JH, Kim K, Park KH, Kim YB. Cell-free DNA Level as a Prognostic Biomarker for Epithelial Ovarian Cancer. *Anticancer Res*. 2012;32(8):3467–71.
- Stroun M, Anker P, Lyautey J, Lederrey C, Maurice PA. Isolation and characterization of DNA from the plasma of cancer patients.

- Eur J Cancer Clin Oncol. 1987;23(6):707–12. doi:10.1016/0277-5379(87)90266-5.
- Holmgren L, Reilly MS, Folkman J. Dormancy of micrometastases: balanced proliferation and apoptosis in the presence of angiogenesis suppression. *Nat Med.* 1995;1(2):149–53. doi:10.1038/nm0295-149.
- Wikman H, Vessella R, Pantel K. Cancer micrometastasis and tumour dormancy. APMIS. 2008;116(7-8):754–70. doi:10.1111/j.1600-0463.2008.01033.x.
- Nicola MH, Bizon R, Machado J, Sollero T, Rodarte RS, Nobre JS, et al. Breast cancer micrometastases: different interactions of carcinoma cells with normal and cancer patients' bone marrow stromata. Clin Exp Metastasis. 2003;20(5):471–9.
- Bettegowda C, Sausen M, Leary RJ, Kinde I, Wang Y, Agrawal N, et al. Detection of circulating tumor DNA in early- and late-stage human malignancies. Sci Transl Med. 2014;6(224):224. doi:10.1126/scitranslmed.3007094.
- Cheng X, Zhang L, Chen Y, Qing C. Circulating cell-free DNA and circulating tumor cells, the "liquid biopsies" in ovarian cancer. *J Ovarian Res.* 2017;10(1):75. doi:10.1186/s13048-017-0369-5.
- Khati S, Jaiswal R, Singh N. Evaluation of Serum HE4 in Ovarian Tumors, Its Comparison with Serum CA125 and Correlation with Histological Subtypes. *Indian J Gynecol Oncol.* 2018;16(4). doi:10.1007/s40944-018-0232-7.
- Board RE, Wardley AM, Dixon JM, Armstrong AC, Howell S, Renshaw L, et al. Detection of PIK3CA mutations in circulating free DNA in patients with breast cancer. *Breast Cancer Res Treat*. 2010;120(2):461–7. doi:10.1007/s10549-010-0747-9.
- Shao X, He Y, Ji M, Chen X, Qi J, Shi W, et al. Quantitative analysis of cell-free DNA in ovarian cancer. *Oncol Lett.* 2015;10(6):3478–82. doi:10.3892/ol.2015.3771.
- Ignatiadis M, Lee M, Jeffrey SS. Circulating Tumor Cells and Circulating Tumor DNA: Challenges and Opportunities on the Path to Clinical Utility. Clin Cancer Res. 2015;21(21):4786–800. doi:10.1158/1078-0432.CCR-14-1190.

Author biography

Pravesh Gupta, Junior Resident

Riddhi Jaiswal, Additional Professor

Piyush Tripathi, Research Officer

Vimala Venkatesh, Professor

Cite this article: Gupta P, Jaiswal R, Tripathi P, Venkatesh V. Observational study on serum markers and circulating tumor cells in ovarian cancer. *IP Arch Cytol Histopathology Res* 2021;6(3):181-186.