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Review Article

The role of LncRNAs in the development of cataracts

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ABSTRACT

The prevalence of eye diseases worldwide is dramatically increasing and represent a major concern in underdeveloped and developed regions, especially sight threatening diseases. Ocular diseases, previously associated with a higher depression risk, also impose a substantial economic burden on affected families and society, thus the importance of early detection and accurate treatment in order to avoid and prevent blindness should be emphasized. Cataract, a clouding (opacification) in the normally transparent of lens which leads to a decrease in vision, is most commonly due to aging but may also be present at birth and occur due to trauma or radiation exposure. With the increasing population of elderly people and cataract patients in China, the social burden of cataract is a big challenge at present and will continue to be a challenge in the future. Genetics have been shown to play an important role in the occurrence of eye diseases, with the detection of a numbers of specific gene mutations. LncRNAs have emerged as a novel class of regulatory molecules involved in numerous biological processes and complicated diseases, however the proper connections and pathways they may use to influence the susceptibility to developing cataracts have not yet been completely elucidated. In this review, we focus on the LncRNAs characteristics and its regulation, and summarize these results from separate, independent, cataract-related studies in addition to discussing possible pathways by which LncRNAs might contribute to the development of cataract.

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1. Introduction

A cataract is a clouding (opacification) in the normally transparent of lens which leads to a decrease in vision. In healthy unclouded lens, light is able to pass through to the retina, allowing us to see detail. Around 40 year old of age, the proteins in the lens start to break down and clump together, which leads to the development of a cloudy area on the lens.¹ It often develops slowly and can be uni or bilateral, the symptoms include blurry or double vision, faded colors, halos around light, photophobia, and nyctalopia.² Poor vision caused by cataracts may also result in an increased depression risk and adverse events including falls and fractures.^{3,4} Cataracts are most commonly due

to aging but may also be present at birth and occur due to trauma or radiation exposure. Biological aging is the most common cause of cataracts but other <https://en.wikipedia.org/wiki/Cataract> - cite_note-WHOPri-4 risk factors include diabetes, smoking tobacco, prolonged exposure to ultraviolet radiation, skin diseases, injury, infection, smoking, and genetic factors.⁵

Cataracts are the leading cause of reversible blindness and 33% of visual impairment worldwideremains a severe public health challenge worldwide, especially in China.⁶ With the increasing population of elderly people and cataract patients in China, the social burden of cataract is a big challenge at present and will continue to be a challenge in the future.⁷ Therefore, it is critical to explore the potential risk factors for ARC from an epidemiological perspective to determine the mechanism

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for the formation of cataracts and a potential method for cataract prevention. Surgery is needed only if the cataracts are causing problems and generally results in an improved quality of life.⁸ Cataract surgery is not readily available in many countries, which is especially true for women, those living in rural areas, and those who do not know how to read.⁹ It is the cause of approximately 5% of blindness in the United States and nearly 60% of blindness in parts of Africa and South America.⁹ Blindness from cataracts occurs in about 10 to 40 per 100,000 children in the developing world, and 1 to 4 per 100,000 children in the developed world.¹⁰

The transparency of lens tends to deteriorate with ultraviolet radiation, oxidative stress, age and many other toxic factors, eventually resulting in the development of cataract.^{11,12} Increased proteolysis, alteration of cell cycle, DNA damage, changes in growth and differentiation of lens epithelial cells are the main morphological and functional changes occurring during the process of cataract development.¹³ Accumulating evidence reveals that gene expression in the lens epithelium is significantly altered during cataract formation. For instance, metallothionein IIA, osteonectin and adhesion related kinase are up-regulated in cataractous lenses relative to transparent lenses,^{14–16} whereas many ribosomal proteins and protein phosphatase 2A are down-regulated in cataractous lenses relative to transparent lenses.^{17,18} Metallothionein IIA participates in metal binding and detoxification. Osteonectin is a calcium-binding protein, which serves as a key regulator of cell growth. Decreased protein synthesis, a pathological process involved in the development of cataract can be the result of reduced expression of ribosomal proteins.¹¹

The non-coding region of the genome has recently been recognized to possess a crucial functional importance in normal development and physiology and this discovery has focused increasing attention on its potential to contribute towards diverse disease aetiology.¹⁹ LncRNAs are defined as transcripts with a size ranging from 200 to 100 000 nucleotides, structurally resembling mRNA and presenting little to no protein-coding potential and can be classified into several types according to their genomic locations. Although the vast majority of lncRNAs are situated in the nucleus,²⁰ however, a substantial minority (nearly 15%) are present in the cytoplasm.²¹ LncRNAs can be classified as sense or antisense, the former comprising of those that overlap with protein-coding genes, and the latter comprising those that are antisense transcribed to protein-coding genes. If the promoter and transcript are situated in proximity and also in a head-to-head orientated fashion, the lncRNA is then said to be bidirectional.^{22–24} Many studies reported important regulatory roles for lncRNAs in multiple biological processes, including cell lineage commitment, stem cell maintenance, and cellular phenotype differentiation.^{25–27} Transcriptional regulation may be

influenced by lncRNAs via several modes such as decoy, signal, guide and scaffold.²⁸ They might also act as signals in response to multiple stimuli, participate in recruiting corresponding complexes in order to directly or indirectly silence or activate the expression of a gene.^{29–31} In addition, some lncRNAs may affect gene expression through post-transcriptional events, lncRNAs also participate in the modification process post-translation.³² Nonetheless, several lncRNAs have been implicated in various diseases such as neurodegenerative diseases,^{33–36} multiple tumors and cancers,^{37–45} and common ocular diseases such as glaucoma,⁴⁶ and diabetic retinopathy,⁴⁷ among others, the proper connections and pathways they may use to influence the susceptibility to developing cataracts have not yet been completely elucidated.

In this article, we aim to demonstrate the implication of lncRNAs in cataract development by summarizing results from separate, independent studies.

2. II- LncRNAs in cataracts

2.1. $\beta B1$ -crystallin (*CRYBB1*) and $\beta B2$ -crystallin (*CRYBB2*)

Close to half of all cases of congenital cataracts are reportedly caused by genetic mutations,⁴⁸ a dozen genes have successfully been associated with the development of the condition⁴⁹ such as membrane transport protein genes, cytoskeletal protein gene, crystallin genes and transcription factor genes. Crystallins, composed of α , β and γ , represent ocular lens structural proteins, and are necessary for normal lens transparency and refractive power preservation.⁵⁰ Initially thought to be only present in the lens, however their expression has been detected in various tissues.^{51,52} α -crystallins act as small heat-shock proteins; whereas the functions of $\beta\gamma$ -crystallins is not completely elucidated. β - and γ -crystallins share a common core protein structure with two similar domains, each composed of characteristic key-motifs.^{53–55} Characterized as oligomers, β -crystallin family actually comprises basic ($\beta B1$, $\beta B2$, $\beta B3$) and acidic ($\beta A3/A1$, $\beta A2$, $\beta A4$) proteins.⁵⁶ Basic β -crystallins possess both N-terminal and C-terminal extensions, whereas acidic β -crystallins have only N-terminal extensions.⁵⁷

The discovery of multiple mutations mostly in Chinese families suggests an important role played by *CRYBB1* in congenital cataract. *CRYBB1*-Crystallin Knockout was not only linked to the development of nuclear cataract but is also suspected to induce impaired lysosomal cargo clearance and calpain activation. Homozygous *CRYBB1* deletion mutation underlies autosomal recessive congenital cataract whereas missense mutation S228P and nonsense mutation (p.Q223X) have been previously associated with autosomal dominant congenital cataract. Additionally, the *CRYBB1* mutation (c.347T>C), *CRYBB2* mutation (c.355G>A) are novel in patients with congenital

cataract.^{58–61}

β B2-crystallin (CRYBB2) is highly expressed in the postnatal lens cortex and linked to the development of cataracts, hinting at the alteration of gene expression in the lens epithelium during the development of cataract. Reports about the effect of CRYAA and CRYBA1/CRYBA3's downregulation and the upregulation of the receptor tyrosine kinase adhesion-related kinase (ARK) in a mouse model of age-related cataracts were made by Sheets et al.⁶² Furthermore, metallothionein-IIA, osteonectin and ARK were also proved to be comparatively upregulated in cataractous lenses.^{14–16} Additionally, genes such as GCS1, and POLR2E were also found to be extensively downregulated in cataractous lens.⁶³ Crystallins, including CRYBB2 in particular, are suspected to be able to primarily act as lens' structural proteins.⁶⁴ The relative CRYBB2 protein expression was demonstrated to change markedly during the first year of life,⁶⁵ suggesting that CRYBB2 serves a contributive function in lens development. Moreover, targeted knockout (KO) of CRYBB2 in mice has been demonstrated to induce age-related⁶⁶ and congenital cataracts;⁶⁷ however, its functional significance is not yet known.

Yin and al. tried to analyze lncRNAs and mRNAs by performing expression profiling on CRYBB2 knockdown-induced cataracts and non-treated mice. Their results showed a total of 329 differentially expressed lncRNAs in CRYBB2 KO group mice lenses (A total of 149 lncRNAs identified to be upregulated, 180 lncRNAs were found downregulated).⁶⁸

2.2. MIAT

MIAT, also known as Gomafu in human or Rncr2 in mouse, is a lncRNA initially identified in 2000 as a susceptibility locus for myocardial infarction patients,⁶⁹ located at 22q12.1, highly expressed in retinal precursor cells and highly conserved in placental mammals.^{70,71} It also shows a deregulation in multiple diseases such as neuroendocrine prostate cancer, non-small-cell lung cancer diseases, diabetic cardiomyopathy and neuropathy, chronic chagas disease, chronic lymphocytic leukemia, schizophrenia, ocular neovascularizations, bone disease and ischemic stroke.^{71–74}

A recent study performed lncRNA microarray on cataractous and transparent lens and identified 38 differentially expressed lncRNAs, including 21 up-regulated and 17 down-regulated among which MIAT's expression level was significantly higher in cataractous lens. Further investigations on MIAT expression level on aqueous humor and whole blood collected from healthy controls, cataract, glaucoma and PVR patients in order to verify their early findings. The results showed that MIAT is significantly up-regulated in the plasma and aqueous humor of cataract patients, but not in other patients with

glaucoma, PVR.^{75,76} Oxidative stress is considered an important risk factor for the development of age-related cataract because lens cells are constantly exposed to reactive oxygen species including hydroxyl radical (\cdot OH), free radicals superoxide ($O_2^{\cdot-}$), H_2O_2 and hypochlorous acid (HClO).^{13,16} Accumulated products of oxidative stress is present in cataract patients and may explain why they possess an higher MIAT expression level compared to the matched controls. The viability and proliferation ability of human lens epithelial cells could be reduced by oxidative stress, whereas they were showed to be further decreased when subjected to a MIAT's knockdown, implying that an increased MIAT level is possibly a response against oxidative stress. Opacification of posterior capsule is the main complication of cataract surgery. Following the insult of surgery, residual lens epithelial cells rapidly grow at the equator and under the anterior lens capsule. These cells proliferate and migrate onto the posterior capsule.^{77,78} The response of lens epithelial cells can be considered a wound healing reaction resulting from the activation of inflammatory cells and production of cytokines and growth factors after surgery.⁷⁹ The above findings suggest that MIAT might potentially be used as a specific biomarker in early detection of cataract.

2.3. lncRNA H19

lncRNA H19, is a transcript from the H19/IGF2 gene, and located at 11p15.5 locus. Its expression is very high in fetus but tends to decrease progressively after birth, and these favorable characteristics enable its function as a genetic biomarker.⁸⁰ It actually regulates various biological processes by acting as a ceRNA that released the miRNA's targets via the competition for miRNA to influence the related protein factors.⁸¹ lncRNA H19 has been found to be implicated via different signaling pathways in many kinds of tumors, and its mutation in mouse zygotes lead to prenatal lethality, suggesting a important role in normal growth and development.^{82–84} and prompting new research paradigm to further understand intrinsic mechanisms of lncRNAs.⁸⁵

MicroRNAs (miRNAs) are the most frequently studied class of the non coding RNAs, and possess a length of ~22 nucleotides (nt). They participate in the mediation of post transcriptional gene silencing via controlling the translation of mRNA into proteins.^{86,87} Specifically expressed in lens, previous researches have tried to decipher the regulating role of miRNAs in HLECs function^{88,89} and found out that in some cases, miRNAs level decreased in cataractous lens cells from rats.⁹⁰ Cheng and al. recently used sequencing technology aiming to identify and compare expression of lncRNAs in age-related cataracts and further explore the oxidative damage repair mechanism. In their experiment, more than 50 lncRNAs were differently expressed, among which lncRNA H19 was up-regulated at early age-related cataract development and ultra violet irradiation-

induced oxidative damage model cells, whereas miRNA 29a expression decreased in the three types of early ARC and in HLECs exposed to UVB irradiation.⁹¹ Reactive oxygen species accumulation oxidative stress by inducing damage to the DNA can lead to the age related cataract development.¹³ human thymine DNA glycosylase plays an important role in aberrant BER pathway of oxidatively damaged DNA. These data indicate that lncRNA H19 could be a useful biomarker of early age-related cataract and deciphering the proper relation between lncRNA H19 and miR-29a may represent a target for age-related cataract treatment.

3. Conclusion

III characteristics and regulation, and tried to discuss potential pathways by which β B1-crystallin (CRYBB1), β B2-crystallin (CRYBB2), MIAT, lncRNA H19 may contribute to the development of cataracts. However, further researches and investigations are needed to discover vast number of cataract-associated lncRNAs, their characteristics and expression patterns, and decipher pathways of their involvement and role in the pathogenesis of this potentially blinding condition. We stress that this article creates a paradigm for future studies of lncRNAs in the early determination and monitoring of the evolution of cataract and may prove to be useful for early determination of whether a patient with a suspected lncRNAs should receive prioritized treatment to help slow the progression of the condition, and subsequently avoid blindness, and retain quality of life.

4. Contributorship Statement

Involved: in conception, design (CY), literature search and writing of the manuscript (CY); supervision (CY), critical reading (CY, DVK), approval of the final proofs of the article (CY, DVK).

5. Competing Interests

The authors have declared that no competing interest exists.

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8. Conflict of Interest

None.

References

1. Thompson J, Lakhani N. Cataracts. *Prim Care*. 2015;42(3):409–23. doi:10.1016/j.pop.2015.05.012.
2. Allen D, Vasavada A. Cataract and surgery for cataract. *BMJ*. 2006;333(7559):128–32. doi:10.1136/bmj.333.7559.128.
3. Gimbel HV, Dardzhikova AA. Consequences of waiting for cataract surgery. *Curr Opin Ophthalmol*. 2011;22(1):28–30.
4. Mccusker S, Koola MM. Association of Ophthalmologic Disorders and Depression in the Elderly: A Review of the Literature. *Prim Care Companion CNS Disord*. 2015;17(4). doi:10.4088/PCC.14r01731.
5. Srinivasan S. Incidence, Progression, and Risk Factors for Cataract in Type 2 Diabetes. *Invest Ophthalmol Vis Sci*. 2017;58(13):5921–9. doi:10.1167/iovs.17-22264.
6. Brian G, Taylor H. Cataract blindness—challenges for the 21st century. *Bull World Health Organ*. 2001;79(3):249–56.
7. Zhou Q. The Epidemiology of Age-Related Eye Diseases in Mainland China. *Ophthalmic Epidemiology*. 2007;14(6):399–407.
8. Lamoureux EL. The impact of cataract surgery on quality of life. *Curr Opin Ophthalmol*. 2011;22(1):19–27.
9. Rao GN, Khanna R, Payal A. The global burden of cataract. *Curr Opin Ophthalmol*. 2011;22(1):4–9. doi:10.1097/ICU.0b013e3283414fc8.
10. Pandey SK. Pediatric cataract surgery and intraocular lens implantation: current techniques, complications, and management. *Int Ophthalmol Clin*. 2001;41(3):175–96.
11. Hejtmancik JF, Kantorow M. Molecular genetics of age-related cataract. *Exp Eye Res*. 2004;79(1):3–9.
12. West SK, Valmadrid CT. Epidemiology of risk factors for age-related cataract. *Surv Ophthalmol*. 1995;39(4):323–34.
13. Michael R, Bron AJ. The ageing lens and cataract: a model of normal and pathological ageing. *Philos Trans R Soc Lond B Biol Sci*. 1568;366(1568):1278–92. doi:10.1098/rstb.2010.0300.
14. Hawse J, Padgaonkar VA, Leverenz VR, Pelliccia SE, Kantorow M, Giblin FJ, et al. The role of metallothionein IIa in defending lens epithelial cells against cadmium and TBHP induced oxidative stress. 2006;12:342–9.
15. Gilmour DT, Lyon GJ, Carlton MB, Sanes JR, Cunningham JM, Anderson JR, et al. Mice deficient for the secreted glycoprotein SPARC/osteonectin/BM40 develop normally but show severe age-onset cataract formation and disruption of the lens. *EMBO J*. 1998;17(7):1860–70. doi:10.1093/emboj/17.7.1860.
16. Hawse JR, Hejtmancik JF, Horwitz J, Kantorow M. Identification and functional clustering of global gene expression differences between age-related cataract and clear human lenses and aged human lenses. *Exp Eye Res*. 2004;79(6):935–40. doi:10.1016/j.exer.2004.04.007.
17. Martens E, Stevens I, Janssens V, Vermeesch J, Götz J, Goris J, et al. Genomic organisation, chromosomal localisation tissue distribution and developmental regulation of the PR61/B' regulatory subunits of protein phosphatase 2A in mice. *J Mol Biol*. 2004;336(4):971–86. doi:10.1016/j.jmb.2003.12.047.
18. Zhang W. Decreased expression of ribosomal proteins in human age-related cataract. *Invest Ophthalmol Vis Sci*. 2002;43(1):198–204.
19. Esteller M. Non-coding RNAs in human disease. *Nat Rev Genet*. 2011;12(12):861–74. doi:10.1038/nrg3074.
20. Zhang K, Shi Z, Chang Y, Hu ZM, Qi HX, Hong W, et al. The ways of action of long non-coding RNAs in cytoplasm and nucleus. *Gene*. 2014;547(1):1–9. doi:10.1016/j.gene.2014.06.043.
21. Rashid F, Shah A, Shan G. Long Non-coding RNAs in the Cytoplasm. *Genomics Proteomics Bioinformatics*. 2016;14(2):73–80. doi:10.1016/j.gpb.2016.03.005.
22. Panzeri I, Rossetti G, Abrignani S, Pagani M. Long Intergenic Non-Coding RNAs: Novel Drivers of Human Lymphocyte Differentiation. *Front Immunol*. 2015;6:175. doi:10.3389/fimmu.2015.00175.
23. Fatica A, Bozzoni I. Long non-coding RNAs: new players in cell differentiation and development. *Nat Rev Genet*. 2014;15(1):7–21.
24. Iltot NE, Ponting CP. Predicting long non-coding RNAs using RNA sequencing. *Methods*. 2013;63(1):50–9.
25. Guttman M. lincRNAs act in the circuitry controlling pluripotency and differentiation. *Nat*. 2011;477(7364):295–300.

26. Rapicavoli NA, Blackshaw SE, Blackshaw S. The long noncoding RNA RNCR2 directs mouse retinal cell specification. *BMC Dev Biol.* 2010;10:49. doi:10.1186/1471-213X-10-49.
27. Ramos AD. Integration of genome-wide approaches identifies lncRNAs of adult neural stem cells and their progeny in vivo. *Cell Stem Cell.* 2013;12(5):616–28.
28. Fang Y, Fullwood MJ. Roles, Functions, and Mechanisms of Long Non-coding RNAs in Cancer. *Genomics Proteomics Bioinformatics.* 2016;14(1):42–54. doi:10.1016/j.gpb.2015.09.006.
29. Takahashi K. Long noncoding RNA in liver diseases. *Hepatology.* 2014;60(2):744–53.
30. Mercer TR, Mattick JS. Structure and function of long noncoding RNAs in epigenetic regulation. *Nat Struct Mol Biol.* 2013;20(3):300–7.
31. Chu C. Genomic maps of long noncoding RNA occupancy reveal principles of RNA-chromatin interactions. *Mol Cell.* 2011;44(4):667–78.
32. Huarte M. The emerging role of lncRNAs in cancer. *Nat Med.* 2015;21(11):1253–61.
33. Tan L. Non-coding RNAs in Alzheimer's disease. *Mol Neurobiol.* 2013;47(1):382–93.
34. Lourenco GF. Long noncoding RNAs in TDP-43 and FUS/TLS-related frontotemporal lobar degeneration (FTLD). *Neurobiol Dis.* 2015;82:445–54. doi:10.1016/j.nbd.2015.07.011.
35. Johnson R. Long non-coding RNAs in Huntington's disease neurodegeneration. *Neurobiol Dis.* 2012;46(2):245–54.
36. Wu P, Zuo X, Deng H, Liu X, Liu L, Aimin J. Roles of long noncoding RNAs in brain development, functional diversification and neurodegenerative diseases. *Brain Res Bull.* 2013;97:69–80. doi:10.1016/j.brainresbull.2013.06.001.
37. Kumar M, Devaux RS, Herschkowitz JI. Molecular and Cellular Changes in Breast Cancer and New Roles of lncRNAs in Breast Cancer Initiation and Progression. *Prog Mol Biol Transl Sci.* 2016;144:563–86. doi:10.1016/bs.pmbts.2016.09.011.
38. Zhang Q. The complexity of bladder cancer: long noncoding RNAs are on the stage. *Mol Cancer.* 2013;12(1):101. doi:10.1186/1476-4598-12-101.
39. Zhao J. Long noncoding RNAs in liver cancer: what we know in 2014. *Expert Opin Ther Targets.* 2014;18(10):1207–18. doi:10.1517/14728222.2014.941285.
40. Chen J, Wang R, Zhang K, Chen LB. Long non-coding RNAs in non-small cell lung cancer as biomarkers and therapeutic targets. *J Cell Mol Med.* 2014;18(12):2425–36.
41. Huang X. LncRNAs in pancreatic cancer. *Oncotarget.* 2016;7(35):57379–90.
42. Zhou S, Wang J, Zhang Z. An emerging understanding of long noncoding RNAs in kidney cancer. *J Cancer Res Clin Oncol.* 2014;140(12):1989–95. doi:10.1007/s00432-014-1699-y.
43. Zhao M. Long non-coding RNAs involved in gynecological cancer. *Int J Gynecol Cancer.* 2014;24(7):1140–5.
44. Li X, Cao Y, Gong X, Li H. Long noncoding RNAs in head and neck cancer. *Oncotarget.* 2017;8(6):10726–40.
45. Plass CWMF, Gerhauser C, Gerhauser C. Role of lncRNAs in prostate cancer development and progression. *Biol Chem.* 2014;395(11):1275–90. doi:10.1515/hsz-2014-0201.
46. Cissé Y, Bai L, Meng T. LncRNAs in genetic basis of glaucoma. *BMJ Open Ophthalmology.* 2018;3(1):e000131. doi:10.1136/bmjophth-2017-000131.
47. Yan B, Tao ZF, Xiu-Miao L, Zhang H, Yao J, Jiang Q, et al. Aberrant expression of long noncoding RNAs in early diabetic retinopathy. *Invest Ophthalmol Vis Sci.* 2014;55(2):941–51. doi:10.1167/iov.13-13221.
48. Gao X, Cheng J, Lu C, Li X, Li F, Liu C, et al. A Novel Mutation in the Connexin 50 Gene (GJA8) Associated with Autosomal Dominant Congenital Cataract in a Chinese Family. *Curr Eye Res.* 2010;35(7):597–604. doi:10.3109/02713681003725831.
49. Chen C, Sun Q, Gu M, Liu K, Sun Y, Xu X, et al. A novel Cx50 (GJA8) p.H277Y mutation associated with autosomal dominant congenital cataract identified with targeted next-generation sequencing. *Graefes Arch Clin Exp Ophthalmol.* 2015;253(6):915–24. doi:10.1007/s00417-015-3019-x.
50. Tardieu ADMF, Tardieu A. Short-range order of crystallin proteins accounts for eye lens transparency. *Nat.* 1983;302(5907):415–7.
51. Clayton RM, Jeanny JC, Bower DJ, Errington LH. The presence of extralenticular crystallins and its relationship with transdifferentiation to lens. *Curr Top Dev Biol.* 1986;20:137–51. doi:10.1016/s0070-2153(08)60660-2.
52. Xi J, Farjo R, Yoshida S, Kern TS, Swaroop A, Andley UP. A comprehensive analysis of the expression of crystallins in mouse retina. *Mol Vis.* 2003;9:410–9.
53. Blundell T, Lindley PF. The molecular structure and stability of the eye lens: x-ray analysis of gamma-crystallin II. *Nature.* 1981;289(5800):771–7. doi:10.1038/289771a0.
54. Berbers GA, Hoekman WA, Bloemendal H, de Jong W, Kleinschmidt T, Braunitzer G, et al. Homology between the primary structures of the major bovine beta-crystallin chains. *Eur J Biochem.* 1984;139(3):467–79. doi:10.1111/j.1432-1033.1984.tb08029.x.
55. Lampi K, Z Ma MS, Shearer T, Smith J, Smith D, David L, et al. Sequence analysis of betaA3, betaB3, and betaA4 crystallins completes the identification of the major proteins in young human lens. *J Biol Chem.* 1997;272(4):2268–75. doi:10.1074/jbc.272.4.2268.
56. Herbrink P, Van Westreenen HF, Van Westreenen H, Fau - Bloemendal H, Bloemendal H. Further studies on the polypeptide chains of beta-crystallin. *Exp Eye Res.* 1975;20(6):541–8.
57. Jobby MK, Sharma Y. Calcium-binding to lens betaB2- and betaA3-crystallins suggests that all beta-crystallins are calcium-binding proteins. *FEBS J.* 2007;274(16):4135–47. doi:10.1111/j.1742-4658.2007.05941.x.
58. Yang J, Zhu Y, Gu F, He X, Cao Z, Li X, et al. A novel nonsense mutation in CRYBB1 associated with autosomal dominant congenital cataract. *Mol Vis.* 2008;14:727–31.
59. Chen P, Chen H, Xiao-Jing P, Su-Zhen T, Yu-Jun X, Zhang H, et al. Novel mutations in CRYBB1/CRYBB2 identified by targeted exome sequencing in Chinese families with congenital cataract. *Int J Ophthalmol.* 2018;11(10):1577–82. doi:10.18240/ijo.2018.10.01.
60. Hegde S, Kesterson RA, Srivastava OP. CRYbetaA3/A1-Crystallin Knockout Develops Nuclear Cataract and Causes Impaired Lysosomal Cargo Clearance and Calpain Activation. *Chin Med J (Engl).* 2007;120(9):820–4.
61. Hegde S, Kesterson RA, Srivastava OP. CRYbetaA3/A1-Crystallin Knockout Develops Nuclear Cataract and Causes Impaired Lysosomal Cargo Clearance and Calpain Activation. *Invest Ophthalmol Vis Sci.* 2007;48(5):2208–13.
62. Sheets NL. Cataract- and lens-specific upregulation of ARK receptor tyrosine kinase in Emory mouse cataract. *Invest Ophthalmol Vis Sci.* 2002;43(6):1870–5.
63. Ruotolo R, Grassi F, Percudani R, Rivetti C, Martorana D, Maraini G, et al. Gene expression profiling in human age-related nuclear cataract. *Mol Vis.* 2003;9:538–48.
64. Pauli S, Söker T, Klopp N, Illig T, Engel W, Graw J, et al. Mutation analysis in a German family identified a new cataract-causing allele in the CRYBB2 gene. *Mol Vis.* 2007;13:962–7.
65. Lou D, Tong JP, Zhang LY, Chiang SWY, Lam DSC, Pang CP, et al. A novel mutation in CRYBB2 responsible for inherited coronary cataract. *Eye (Lond).* 2009;23(5):1213–20. doi:10.1038/eye.2008.222.
66. Zhang J, Li J, Huang C, Xue L, Peng Y, Fu Q, et al. Targeted knockout of the mouse betaB2-crystallin gene (Crybb2) induces age-related cataract. *Invest Ophthalmol Vis Sci.* 2008;49(12):5476–83.
67. Mothobi ME, Guo S, Liu Y, Chen Q, Yussuf AS, Zhu X, et al. Mutation analysis of congenital cataract in a Basotho family identified a new missense allele in CRYBB2. *Mol Vis.* 2009;15:1470–5.
68. Jia Y, Xiong K, Ren HX, Li W. Identification of long non-coding RNA and mRNA expression in betaBeta2-crystallin knockout mice. *Exp Ther Med.* 2018;15(5):4277–83. doi:10.3892/etm.2018.5949.
69. Ohnishi Y, Tanaka T, Yamada R, Suematsu K, Minami M, Fujii K, et al. Identification of 187 single nucleotide polymorphisms (SNPs) among 41 candidate genes for ischemic heart disease in the Japanese population. *Hum Genet.* 2000;106(3):288–92.

- doi:10.1007/s004390051039.
70. Tsuiji H, Yoshimoto R, Hasegawa Y, Furuno M, Yoshida M, Nakagawa S, et al. Competition between a noncoding exon and introns: Gomafu contains tandem UACUAAC repeats and associates with splicing factor-1. *Genes Cells*. 2011;16(5):479–90. doi:10.1111/j.1365-2443.2011.01502.x.
 71. Ishii N, Ozaki K, Sato H, Mizuno H, Saito S, Takahashi A, et al. Identification of a novel non-coding RNA, MIAT, that confers risk of myocardial infarction. *J Hum Genet*. 2006;51(12):1087–99.
 72. Sone M, Hayashi T, Tarui H, Agata K, Takeichi M, Nakagawa S, et al. The mRNA-like noncoding RNA Gomafu constitutes a novel nuclear domain in a subset of neurons. *J Cell Sci*. 2007;120(15):2498–506. doi:10.1242/jcs.009357.
 73. Blackshaw S, Harpavat S, Trimarchi J, Cai L, Huang H, Kuo WP, et al. Genomic analysis of mouse retinal development. *PLoS Biol*. 2004;2(9):E247. doi:10.1371/journal.pbio.0020247.
 74. Bai Y, Wang W, Zhang Y, Zhang F, Zhang H. lncRNA MIAT suppression alleviates corneal angiogenesis through regulating miR-1246/ACE. *Cell Cycle*. 2019;18(6-7):661–9. doi:10.1080/15384101.2019.1578143.
 75. Shen Y, Dong LF, Zhou RM, Yao J, Song Y, Yang H, et al. Role of long non-coding RNA MIAT in proliferation, apoptosis and migration of lens epithelial cells: a clinical and in vitro study. *J Cell Mol Med*. 2016;20(3):537–48. doi:10.1111/jcmm.12755.
 76. Selin JZ, Lindblad BE, Rautiainen S, Michaëlsson K, Morgenstern R, Bottai M, et al. Are increased levels of systemic oxidative stress and inflammation associated with age-related cataract? *Antioxid Redox Signal*. 2014;21(5):700–4. doi:10.1089/ars.2014.5853.
 77. Wang Y, Li W, Zang X, Chen N, Liu T, Tsonis PA, et al. MicroRNA-204-5p regulates epithelial-to-mesenchymal transition during human posterior capsule opacification by targeting SMAD4. *Invest Ophthalmol Vis Sci*. 2013;54(1):323–32. doi:10.1167/iovs.12-10904.
 78. Mamuya FA, Wang Y, Roop VH, Scheiblin DA, Zajac JC, Duncan MK, et al. The roles of alphaV integrins in lens EMT and posterior capsular opacification. *J Cell Mol Med*. 2014;18(4):656–70. doi:10.1111/jcmm.12213.
 79. Xu H, Chen M, Forrester JV, Lois N. Cataract surgery induces retinal pro-inflammatory gene expression and protein secretion. *Invest Ophthalmol Vis Sci*. 2011;52(1):249–55. doi:10.1167/iovs.10-6001.
 80. Pachnis V, Belayew A, Tilghman SM. Locus unlinked to alpha-fetoprotein under the control of the murine raf and Rif genes. *Proc Natl Acad Sci*. 1984;81(17):5523–7. doi:10.1073/pnas.81.17.5523.
 81. Cai X, Cullen BR. The imprinted H19 noncoding RNA is a primary microRNA precursor. *RNA*. 2007;13(3):313–6. doi:10.1261/rna.351707.
 82. Brunkow ME, Tilghman SM. Ectopic expression of the H19 gene in mice causes prenatal lethality. *Genes Dev*. 1991;5(6):1092–101.
 83. Li CF, Li YC, Wang Y, Sun LB. The Effect of LncRNA H19/miR-194-5p Axis on the Epithelial-Mesenchymal Transition of Colorectal Adenocarcinoma. *Cell Physiol Biochem*. 2018;50(1):196–213. doi:10.1159/000493968.
 84. Ismail DM, Shaker OG, Kandeil MA, Hussein RM. Gene Expression of the Circulating Long Noncoding RNA H19 and HOTAIR in Egyptian Colorectal Cancer Patients. *Genet Test Mol Biomarkers*. 2019;23(9):671–80. doi:10.1089/gtmb.2019.0066.
 85. Salmena L, Poliseno L, Tay Y, Kats L, Pandolfi PP. A ceRNA hypothesis: the Rosetta Stone of a hidden RNA language? *Cell*. 2011;146(3):353–8. doi:10.1016/j.cell.2011.07.014.
 86. Mendell JT. MicroRNAs: critical regulators of development, cellular physiology and malignancy. *Cell Cycle*. 2005;4(9):1179–84.
 87. He L, Hannon GJ. MicroRNAs: small RNAs with a big role in gene regulation. *Nat Rev Genet*. 2004;5(7):522–31.
 88. Chien KH, Chen SJ, Liu JH, Chang HM, Woung LC, Liang CM, et al. Correlation between microRNA-34a levels and lens opacity severity in age-related cataracts. *Eye (Lond)*. 2013;27(7):883–8. doi:10.1038/eye.2013.90.
 89. Zhang F, Meng W, Tong B. Down-Regulation of MicroRNA-133b Suppresses Apoptosis of Lens Epithelial Cell by Up-Regulating BCL2L2 in Age-Related Cataracts. *Med Sci Monit*. 2016;22:4139–45. doi:10.12659/msm.896975.
 90. Kubo E, Hasanova N, Sasaki H, Singh DP. Dynamic and differential regulation in the microRNA expression in the developing and mature cataractous rat lens. *J Cell Mol Med*. 2013;17(9):1146–59. doi:10.1111/jcmm.12094.
 91. Cheng T. lncRNA H19 contributes to oxidative damage repair in the early age-related cataract by regulating miR-29a/TDG axis. *J Cell Mol Med*. 2019;23(9):6131–9.

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