

RESEARCH ARTICLE

Process Variables Combination of Roasted African Breadfruit Seed Flour and the Essential Amino Acid Needs of Infants and Children

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ABSTRACT

Infants and growing children require eight essential amino acids for healthy growth and well-being. The amino acids must be at optimum values to satisfy their metabolic requirements. Breadfruit (*v. Decne*) seed is a widely consumed legume in tropics and subtropical regions of the world. In Nigeria, it is staple and an important source of dietary nutrients for adult, infants, and children. The study aimed to identify the roasting conditions of temperature and time at 500 g seed quantity that would yield the optimum contents of eight essential amino acids in flour incorporated into the diets of infants and growing children for the alleviation of malnutrition among children. Experimental roasting used the factorial design. The essential amino acids content of breadfruit seed flour of different treatment conditions was determined using Technicon sequential multisample acid analysis. Results showed that both raw and processed flour contained amino acids essential for infants and growing children. Analysis of amino acids showed retention of the eight essential amino acids for infant and children. Roasting temperature and roasting time had significant ($P < 0.05$) reductive effect on evaluated amino acids. Summation of the evaluated amino acids showed average ranges of 22.56–25.61 g/16 gN, 17.14–23.86 g/16 gN, and 18.56–19.90 g/16 gN, respectively, for flour samples of seeds roasted at 140°C, 160°C, and 180°C at 35, 40, and 45 min. The low values of some essential amino acids evaluated indicated the detrimental effect of higher temperature processing on African breadfruit seeds. Processing African breadfruit seeds into at 140°C and 160°C for 35–45 min would flour of at least 76.92% of recommended dietary allowance human. Results identified 140°C-40 min-500 g as the best process variables combination for an adequate supply of essential amino acids in infant and children diets. Roasting severely depleted the content of tryptophan of the evaluated flour samples. The flour of breadfruit (*v. Decne*) seeds is a good source of amino acids supplement for infants and growing children which should be introduced to mothers in developing nations.

Key words: Breadfruits, essential amino acids, infant and children diets, roasting condition

INTRODUCTION

Malnutrition of infants and children is a serious challenge to the realization of sustainable development goals (SDG) for maternal and child health in developing nations. Low income, illiteracy

insurgency, food, insecurity, infection, and diseases are some of the factors responsible for global nutritional challenges.

In a reduced availability of animal protein due to disruption in supplies with increase in prices beyond the reach of low-income earners, children are faced with limited supply or essential amino acids needed for healthy growth and well-being. To ameliorate the low intake of essential amino acids due to scarcity of animal protein, some protein-rich

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seeds of leguminous plants are added as a substitute for animal protein in the food fed to infants and growing children (Ugwu and Ekwu, 1996). Eight essential amino acids needed by infants and children are histidine, leucine, isoleucine, lysine, methionine, threonine, phenylalanine, and valine. Reports in literature point to the influence of heating temperature and time on the nutritive value of proteins.

Heating of protein foods leads to the decomposition of amino acids, especially valine, leucine, isoleucine, methionine, histidine, etc. The decomposition due to decarboxylation and deamination produces harmful products and reduces biological values of protein which could be responsible for decreased growth of animals fed with more severely heated protein diets reported. Similarly, Due to excessive cooking leads to loss of biological value of proteins.

The growing preference for roasted breadfruit (*v. Decne*) seed flour over soybean flour for complementation of infant and children foods is evident by the preferential choice of mothers for African breadfruit seed flour. Soybean flour has an objectionable beany taste to infants. The flour of breadfruit seed is produced from roasted, milled, and sieved seeds. The flour is mixed as supplements into diets fed to infants and children for the supply of protein, energy, and other nutrients. Breadfruits seed is an important source of essential amino acids (Okaka and Okaka, 2005). When properly processed the flour of breadfruit seeds can furnish the recommended daily allowances of essential amino acids for human beings.

However, the nutritional importance of breadfruit seed flour is challenged by inappropriate roast processing of African breadfruit seeds by illiterate mothers. Growth test on young rats showed a positive correlation between decreased nutritive valued and roasting (temperature-time) effects on diets. To effectively use breadfruit seed flour, as alternative to animal protein good processing is essential. To achieve this objective, mothers need to be taught the appropriate processing and roasting condition needed to produce flour of excellent amino acid contents. The optimization model developed by Nwabueze (2009) for heat treatment of African breadfruits is too professional and technologically sophisticated for domestic applications. There is the need for a simple and practicable template for heat processing

of African breadfruits. The dearth of information in literature on the effect of roasting on the essential amino acids of African breadfruit seeds flour and the nutritional importance of these essential amino acids for infants and children underscore the importance of this study. The result of this study is intended to aid the realization of the nutritional aspects of SDG in relation to maternal and child health of developing nation, through the exploitation of affordable indigenous and yet nutritious legumes.^[1-20]

MATERIALS AND METHODS

Sample collection and pre-treatment of samples

Breadfruit (*v. Decne*) dry seeds were purchased from Ubani market, Umuahia, Nigeria. The seeds were screened for contaminants such as stones and soils and stored in plastic bowls. The screened seeds were washed and air-dried under shade at ambient temperature (28/30°C).

Pre-treatment roasting, dehulling, and evaluation of 500 g mass of cleaned seeds using an electric oven and locally designed dehuller identified the ideal roasting points to be 140°C and 40 min for roasting temperature, time, dehulling yield, and condition of the dehulled endosperm.

Treatment of sample

The Study design employed three factors by three levels experimental layout. The optimization model of Umezuruike *et al.* (2016) and parametric values in literature for roasting of African breadfruit seeds were employed for allocation of values to roasting temperature and time at a mass of 500 g. For ease of use by local processors, three standout points used were 140°C, 160°C, and 180°C and 35 min, 40 min, and 45 min for roasting temperature and time, respectively. The cleansed breadfruit seeds were divided into groups (500 g) and roasted experimentally at 140°C, 160°C, and 180°C for 35 min, 40 min, and 45 min in oven (Fisher Scientific Oven 655, Fisher Scientific Company, USA). The roasted seeds were cooled, dehulled using locally designed dehuller, and milled with a hand mill (Corona Model, Landers/CIA SA) and sieved into flour using 2.0 mm sieves. The samples were appropriately labeled and used for essential amino acid assay. The control (raw seeds)

was not roasted but shade dried at 28⁺-2°C for 48 h, dehulled, milled, and sieved into flour with 2.0 mm sieve.

Determination of amino acid profile

The amino acid profile of raw and roasted breadfruit flour was determined using the method described by Spackman *et al.* (1958). The samples were dried to constant weights in oven, defatted, hydrolyzed, and then evaporated using rotary evaporator. The evaporated samples were loaded into amino acid analyzer (Technicon sequential multisample [TSM]). Thirty milligram of each sample was mixed with 7 ml of NHCL in a glass ampoule. Oxygen was expelled from the mixture by passing nitrogen into the ampoule. The glass ampoule was heat sealed using a Bunsen burner flame and placed in an oven (105°C) for 22 h. After heating, the glass ampoules were allowed to cool, the tip opened and the content filtered. The filtrate was evaporated to dryness using rotary evaporator at 40°C.

The residue was dissolved in 5 ml acetate buffer (pH 2.0) and stored in refrigerator using plastic bottle. 5–10 µL of each sample was placed in amino acid

cartridges and loaded into the amino acid analyzer. The TSM analyzer separates and analyzes amino acids into acidic, basic, and neutral amino acids. The data generated from the Technicon sequential multisample analyzer were quantitatively determined against the standard Technicon autoanalyzer chart (No. 011-648-0; Technicon Instruments, Tarrytown New York, USA).^[21-30]

Interpretation of results

For statistical evaluation, data of study were analyzed using Minitab statistical software version 15 of Minitab Inc., Pen., USA. Analysis of variance and characterization of variables at 95% confidence limit were used to describe the level of differences among the test samples at mid and extreme roasting temperatures and roasting time.

RESULTS AND DISCUSSION

Amino acid content of samples

The essential amino acid compositions of the processed breadfruitseed flour samples are presented in Tables 1-3.

Table 1: Amino acid content (g/16 gN) of flour of seeds roasted at 140°C

Amino acids	35 min	40 min	45 min	Standard deviation	Standard error of mean
Histidine	2.20	2.23	2.20	0.5831	0.3366
Isoleucine	3.01	2.90	2.89	0.5578	0.3219
Leucine	6.00	5.70	5.01	0.4670	0.2696
Lysine	4.27	4.18	3.11	0.6137	0.3543
Methionine	0.75	0.73	0.70	0.0416	0.0240
Phenylalanine	4.05	3.99	3.80	0.1887	0.1090
Threonine	2.73	2.70	2.75	0.2203	0.1271
Value	2.41	3.18	2.1	0.2157	0.1245
Covariance	0.993	0.978	0.972		

Table 2: Amino acid content (g/16 gN) of flour of seeds roasted at 160°C

Amino acids	35 min	40 min	45 min	Standard deviation	Standard error of mean
Histidine	2.30	2.20	1.19	0.3056	0.1766
Isoleucine	3.11**	2.28	2.05	0.6658	0.0384
Leucine	4.53**	3.99	3.60	0.5076	0.2930
Lysine	3.64	3.49	2.51**	0.6453	0.3725
Methionine	0.69	0.63	0.61	0.0251	0.0145
Phenylalanine	3.47	3.35	3.10	0.1305	0.0753
Threonine	2.71	2.31	2.35	0.2516	0.0145
Value	2.41	2.01	2.07	0.5560	0.3211
Covariance	0.990	0.985	0.982		

Table 3: Amino acid content (g/16 gN) of flour of seeds roasted at 180°C

Amino acids	35 min	40 min	45 min	Standard deviation	Standard error of mean
Histidine	1.80	1.50	1.10	0.3511	0.2028
Isoleucine	1.70	2.38	2.30	0.3716	0.2145
Leucine	4.03	3.39	3.46	0.3510	0.2027
Lysine	3.30	2.91	2.88	0.2343	0.1353
Methionine	0.60	0.57	0.55	0.0251	0.0145
Phenylalanine	3.20	3.39	3.27	0.0960	0.0555
Threonine	2.21	2.01	2.10	0.1001	0.0578
Value	3.05	2.90	2.90	0.0866	0.5000
Covariance	0.941	0.971	0.939		

All essential amino acids in the control are present in the processed flour. These essential amino acids are histidine, isoleucine, leucine, lysine, methionine, phenylalanine, threonine, and valine. The values of all the amino acids analyzed progressively reduced as the roasting temperature was extended from 140°C to 180°C for the duration of 35–45 min. The reported content of the essential amino acids is attributable to roasting condition (different roasting temperature and time). The essential amino acid contents differ significantly in line with applied temperature and duration as consistent with report in literature on the depleting effect of roasting on essential amino acids (Mauron, 1982). Similar report by Adeyeye (2010) showed that roasting of groundnuts resulted in losses in contents for lysine (15.9–27.6%), histidine (4.23–16.5%), threonine (40.1–60.6%), methionine (38.0–63.4%), and isoleucine (13.3–31.8%). Comparatively, the amino acids of African breadfruit seeds are more heat susceptible than groundnuts. The difference in heat liability of the amino acids could be due to differences in the physical dimensions of groundnuts and African breadfruits and heat transfer rates. The physical dimensions of African breadfruit seeds are an important factor of heat processing. The reported maximum loss of essential amino acids occurred at 180°C. Delamination, Millard reactions, and other biochemical reactions have been reported to be responsible for changes in the content of amino acids (Nkatamiya *et al.*, 2007).

Amino acid contents of flour samples produced under different roasting conditions

Results of the amino acid contents of flour samples produced with seeds roasted at 140°C for 35, 40, and 45 min are shown in Table 1.

At 35 min, amino acid contents did not differ significantly except for methionine that reduced (50.77%) significantly. At 35 and 40 min roasting time, the difference in content of amino acids was only 0.74%. Similarly, only slight difference in values was observed for 40 min roasting time. Compared with control, the different roasting time resulted in total amino acid contents of 70.50% (35 min), 71.12% (45 min), and 62.64% (45 min).

Results of the amino acid contents of flour samples produced with seeds roasted at 160°C for 35, 40, and 45 min are presented in Table 2.^[31–38]

Contents of lysine, methionine, and phenylalanine were significantly reduced in flour samples. Observed decreases across roasting time ranged from 7.10 to 2.5 g for lysine, 1.5 to 0.61 g for methionine, and 6.10 to 3.10 g for phenylalanine. Summations of essential amino acid contents showed that flour samples produced at 140°C and 160°C differed by 8.38%. Compared with control, flour samples produced with seeds roasted at 160°C were 66.26%, 56.29%, and 48.46% for 35, 40, and 45 min, respectively.

The process variables combination of 160°C-40 min-500 g marked the start point of significant ($P < 0.05$) decreases in the amino acid contents of roasted African breadfruits.

Roasting at 180°C for 35, 40, and 45 min resulted to significant decreases in amino acid contents [Table 3].

Histidine, isoleucine, lysine, methionine, and phenylalanine showed high heat liability to high temperature roasting. Summations of amino acid contents showed that flour samples produced with seeds roasted at 180°C for 35, 40, and 45 min were 55.26%, 52.62%, and 51.54%, respectively, compared with control.

All roasting variable combinations showed positive correlation ($r > 0.09$) with amino acid contents of the flour samples.

Effects of roasting on the essential amino acids contents of samples

Histidine

Histidine content of processed breadfruit seed flour ranged from 1.9 to 1.29 g/16 gN. Percentage loss in histidine ranged from 26.90% to 65.19%. The effect of roasting temperature and time on histidine was significant ($P < 0.05$). Reductive effect of temperature was linear to mid-temperature (160°C) then squared at higher temperature, reaching the peak loss of 65.19% at 180°C. Less than 50% of histidine was lost at 160°C.

The relationship between roasting variables combination with histidine was described as follows:

$$\text{Histidine} = -16.8033 + 0.1538RT - 0.005RT^2 \text{-----}$$

$$\text{-----R}^\circ = 95.56\% \text{-----(1)}$$

Isoleucine and leucine

Isoleucine and leucine make up one-third of the human muscle (Renie, 2015) which underscores their importance in the nutrition of infants and growing children. As part of hemoglobin, isoleucine and leucine are needed for hormonal synthesis and energy generation in cells and tissues. The isoleucine content of the processed flour ranged from 2.050 g to 3.11 g/16 gN. Percentage loss was between 13.61% and 63.88% with an average loss of 35.27% at mid-temperature (160°C) roasting conditions. Loss of isoleucine was comparable with histidine. The effect of roasting on isoleucine contents of the flour samples was significant ($P < 0.05$). Unlike isoleucine which showed heat stability at lower to mid-temperature, leucine exhibited rapid losses at lower (140°C) to mid (160°C) temperatures. From an initial value of 6.80 g/16 gN (control), leucine loss of leucine in the processed flour ranged from 32.35 to 47.06% at mid-temperature (160°C) to 58.97% at 180°C. The loss of leucine of that value is not nutritionally important. Although leucine showed heat stability at 140–160°C, the effect of roasting temperature was significant.

The effects of roasting variables combinations on isoleucine and leucine were described as follows:

$$\text{Isoleucine} = 26.8837 - 0.6990RT + 0.0010RTRM \text{--}$$

$$\text{----R}^\circ = 83.05 \text{-----(2)}$$

$$\text{Leucine} = 62.3514 - 1.5151RT + 0.0185RT \text{-----}$$

$$\text{---R}^\circ = 70.51 \text{-----(3)}$$

Lysine

Lysine showed higher temperature sensitivity compared with other essential amino acids of the processed flour. From an initial lysine content of 7.10 g/16 gN (control), the pattern of losses of lysine was reflective of temperature-time relationship. Roasting temperature effect on lysine was significant and described by,

$$\text{Lysine} = 5.65976 - 0.007RT \text{-----}$$

$$\text{-----R}^\circ = 90.08 \text{-----(4)}$$

Methionine

Methionine values in processed breadfruit seed flour ranged from 0.55 to 0.75 g/16 gN. Methionine showed a rapid significant ($P < 0.05$) loss (57.52%) in content. Comparatively, methionine is more stable to roasting temperature than lysine. Compared with other essential amino acids, methionine suffered the highest (57.52%) loss at mid-point (160°C) temperature. Roasting time was a significant term and its effect on methionine under the experimental conditions was significant. The effect of roasting time on methionine could be explained by the following equation:

$$\text{Methionine} = 2.3325 + 0.0052RT \text{-----}$$

$$\text{----R}^\circ = (93.39) \text{----(5)}$$

Phenylalanine

Phenylalanine is less stable than methionine during roasting of breadfruit seeds for flour. From initial value of 6.10 g/16 gN (control), phenylalanine losses ranged from 34.43% to 56.72%. Loss of phenylalanine through roasting temperature is comparable with leucine. The rapid depletion observed at temperature (140°C) emphasized the importance of low temperature processing for phenylalanine. The equation describing the effect of roasting time on contents of phenylalanine can be written without the non-significant terms as follows:

$$\text{Phenylalanine} = 19.3587 - 0.0919RT \text{-----}$$

$$\text{-R}^\circ = (78.91) \text{-----(6)}$$

Table 4: Amino acid content (g/16 gN) of flour of seeds roasted at 160°C

Amino acids	Control	Tolerable g/day	FAO/WHO (1982) g/day	FAO/WHO (1991) g/day
Histidine	3.16	-	-	2.50
Isoleucine	3.60	4.5	-	4.80
Leucine	6.80	5.6	4.0	4.90
Lysine	7.10	14	7.0	4.30
Methionine	1.53	5.8	5.5	2.30
Phenylalanine	6.10	-	3.5	2.2-4.1
Threonine	3.22	6.2	4.0	2.90
Value	4.50	-	5.0	4.60

Threonine

Threonine exhibited heat stability up to 180°C process variables conditions. Losses of threonine in the processed flour ranged from 25.37% to 30.81%. Temperature was a significant term on threonine retention in roasted African breadfruit seed flour. The significant relationship could be explained using the equation.

$$\text{Threonine} = 6.4176 + 0.0055\text{RM}^\circ \text{-----}$$

$$-R^\circ = (84.88) \text{-----} (7).$$

Valine

Valine is an essential amino acid needed by infants and children for good metabolism energy generation, mental alertness, and calm disposition (Lehningers *et al.*, 2000). Losses in the valine contents of the roasted seeds flour peaked at 54.89% (180°C). The squared effect of roasting temperature on valine was directly proportional to temperature up to 160°C and mathematically described as follows:

$$\text{Valine} = 24.9539 + 0.0851\text{RM}^\circ \text{-----}$$

$$-R^\circ = (59.07) \text{-----} (8).$$

Essential amino acids needs of infants and children

The tolerable values and recommended dietary allowances (RDAs) for essential amino acids for infants and children are shown in Table 4.

Histidine, a component of hemoglobin is needed by infants and growing children for growth, tissue maintenance, formation of antibodies, and good reproductive development (Francis, 1986; Enwere, 1986).

Low histidine in infant diet places extra demand on leucine, tryptophan, and valine (Passmore, 1981), leading to nitrogen imbalance and adverse rates

of growth and metabolism. Processed breadfruit seed flour can furnish the RDA (20–30 mg/kg) requirement for histidine (FAO/WHO, 1991). However, it is important to note that roasting of African breadfruit seeds used for flour must not exceed 160°C at 30–40 min. Processing above 160°C would result to severe loss and pressure on leucine, tryptophan, and valine, whose values are already at threshold in breadfruit (*v. Decne*) seed flour.

Roasted breadfruit seed flour is adequate in meeting RDA of isoleucine (28–83 mg/kg) for infants and children. Results show that roasted breadfruit seed flour used to supplement diet of infants and growing children is high in leucine.

The complementary role of isoleucine and leucine in diet reduces the adverse consequences of their deficiency (Imura and Okada, 1998). Mental and physical disorders in children are common consequences of such deficiency. Roasting of breadfruit seeds used for diet of infants and children is detrimental to its isoleucine contents. Roasted breadfruit seed flour is a major source of leucine for infants and growing children.

Loss of lysine is of nutritional importance for infants as it is needed for absorption of calcium and formation of collagen. Lysine deficiency in infant diet could lead to poor bone formation, poor skin integrity, poor immunity, and certain childhood diseases.

Although the lysine contents are within RDA (22–99 mg/kg) requirement, processing breadfruit seeds into flour for infant and children diets at above 140°C had adverse effect on the yield of lysine. The abundance of lysine in legumes makes legume-cereal complementation an ideal nitrogen balance method in diets. However, in regions without adequate supply of legumes, lysine

deficiency is an important concern over limitations against healthy bone formation and healthy skin in human.

Methionine is needed for thiamine synthesis and important for infant and children. Its deficiency will result to hyperactivity and poor skin, nail, and hair formation in children. Methionine and lysine maintain a synergy in human body. It is essential for absorption of calcium, collagen formation, nitrogen balance, fat metabolism, anti-oxidation, and proper development of the immune system. Deficiency in methionine or lysine results to inter-methionine-lysine pressure and nitrogen imbalance in the amino acid pool of infants, especially when the diet is deficient in threonine. The RDA (22–49 mg/kg) of methionine for infants and children is marginally met as observed in test flours. Although methionine can be synthesized from cysteine, a critical draw on cysteine leads to excessive draw on tryptophan. As both cysteine and tryptophan are limiting in breadfruits, it is important that processing temperature must not exceed 140°C for 40 min to preserve methionine in the diet of infants and growing children.

Roasted breadfruit seed flour can furnish the RDA (22–141 mg/kg) of phenylalanine for infants and children. As an important constituent of thyroid hormones, phenylalanine is needed by infants and children for learning, vitality, and alertness. The phenylalanine content of African breadfruit seed flour suggested the complementation of breadfruit seed flour with soybean flour is not necessary in infant and children diets.

Threonine is responsible for bone formation, good muscle, good skill, and sugar metabolism in children. The content of threonine in roasted (120–180°C) breadfruit seed flour can furnish the RDA (2.90 g/day) requirement of threonine for infants and children (FAO/WHO, 1991). However, any deficiency can be ameliorated by high lysine content of the flour.

Breadfruit seed flour meets the RDA (98 mg/kg) requirement of valine for infants and children.

CONCLUSION

The effects of roasting on the eight essential amino acids of breadfruit (v. Decne) seed were determined

by comparing the values of amino acids in roasted samples and control.

This study has shown that essential amino acids need of infants and children for good health and well-being can be optimally harnessed from breadfruit seed flour through appropriate roasting condition. Results of the study implied the promise of roasted breadfruit seed flour as an important source for the supply of essential amino acid needs of infants and children. Furthermore, an important means of ameliorating childhood protein-energy malnutrition among children of economic disadvantaged parents and during periods of limited regular animal protein supply due to dislocations of agricultural activities.

REFERENCES

1. Adeyeye ET. Effect of cooking and roasting on the amino acid composition of raw groundnut (*Arachis hypogaea*) seeds. *Acta Sci Pol Tech Aliment* 2010;2:201-16.
2. Anderson SA, Raiten DJ. Safety of Amino-acids Used as Dietary Supplements. FAO Document. Washington: Federated American Societies of Experimental Biology; 1992.
3. Balch P, Balch J. Prescriptions for Nutritional Healing. New York: Avery Books; 2000.
4. Borton B. Human Nutrition. New York: McGraw Hill; 1978.
5. Enwere NJ. Food of Plant Origin. Nsukka: Afro Orbis Publisher; 1998. p. 194-9.
6. Anyalogba EA, Oyeike EN, Monanu MO. Effects of heat treatment on the amino acid profile of *Plukenetia conophora* seeds kernal flours. *Int J Biochem Res Rev* 2015;3:121-31.
7. FAO/WHO Protein Quality Evaluation. Report of a joint FAO/WHO expert Consultation. Food and Agricultural Organization of the United Nation, Rome. Food and Nutrition Paper No 51. 1991.
8. FAO/WHO/UNU Protein and Amino-acid Requirement in Human Nutrition. Geneva: WHO Press; 2007. p. 150.
9. Fau D, Peret J, Hadjiisky P. Effect of ingestion of high protein or excess methionie diets by rats for two-year. *J Nutr* 1998;118:128-33.
10. Food and Nutrition Board/Institute of Medicine. Protein and Amino-Acids: Dietary Reference Intake for Energy. Washington DC: Nat Academy Press; 2002. p. 589-768.
11. Francis D. Nutrition for Children. Oxford: Blackwell; 1986.
12. Harper AE, Benevenga NJ, Wohlheuter RM. Effects of ingestion of disproportionate amount of amino-acids. *Physiol Rev* 1970;50:428-558.
13. Health and Welfare Canada. Report of the Expert Advisory Committee on Amino-acids Catalog No. H42-2/40-Min of Supply and Service Canada. 1990.

14. Imura K, Okada A. Amino-acid metabolism in pediatric patients. *Nutrition* 1998;14:143-8.
15. Iwe MO, Ngoddy PO. Development of mechanical process for the African, breadfruit (*Treculia africana*). *Niger Food J* 2001;19:8-16.
16. Kopple JD, Swendseid ME. Effects of histidine intake on plasma and urine histidine levels nitrogen imbalance in chronically uremic men. *J Nutr* 1988;111:931-42.
17. Lehningers AL, Nelson DL, Cox MM. Principles of Biochemistry. New York: W. H Freeman and Co; 2000.
18. Makinde MA, Arukwe BO, Pettet P. Ukwa seed (*Treculia africana*) protein 1. Chemical evaluation of the protein quality. *J Agric Food Chem* 1985;33:72-4.
19. Mauroa, J. Effect of processing on nutritive value of Food protein. In: Rechcigi M, editor. Handbook of the Nutritional Value of Processed Food. Vol. 1. Florida: CRC Press; 1982. p. 429-72.
20. Meguid MM, Mathews DE, Bier DM, Meredith CN, Soeldner JS, Young VR. Lucine kinetics at graded leucine intakes in young men. *Am J Clin Nutr* 1986;43:770-80.
21. Meredith CN, Wen ZN, Bier DN, Methews DE, Young VR. Valine kinetics at graded value intakes in young men. *Am J Chin Nutr* 1986;43:781-6.
22. Meredith CN, Wen ZN, Bier DM, Mathews DE, Young VR. Lysine kinetic. Graded lysine intakes in young men. *Am J Clin Nutr* 1986;43:787-94.
23. Mindell E. Prescription Alternatives. New York: McGrawHill; 2003.
24. Nkatamiya UU, Madebo AJ, Manji D, Haggai O. Nutrient contents of seeds of some weld plants. *Afr J Biotechnol* 2007;6:1665-9.
25. Nwabueze TU. Kernel extraction and machine efficiency in dehulled parboiled African breadfruit (*T africana*) whole seeds. *J Food Qual* 2009;32:669-83.
26. Nwosu JN, Ubaonu CN, Banigo EO, Uzouah A. The effects of processing on amino acid profile of 'Oze' (bosque qugolensis) seed flow. *Life Sci J* 2008;5:69-74.
27. Okaka JC, Okaka AN. Foods. Composition, Spoilage and Shelf Life Extension. Enugu: Ocjano Academic Publisher; 2005 85-97.
28. Passmore R, Eastwood MA. Human Nutrition and Dietetics. 8th ed. London: Chrchhill Livingstone; 1981.
29. Reeds PJ. Dispensable and indispensable amino-acids for human. *J Nutr* 2000;30:18355-405.
30. Renie C. Why are the Eight Essential Amino Acids Important; 2015 Available from: <http://www.livestrong.com/article/420433>. [Last accessed on 2018 Dec 03].
31. Snyderman SE, Norton PM, Fowler DI, Holt LE. The essential amino-acid requirement of infants. *Lysine Am J Dis Child* 1959;97:175-85.
32. Solomon D. Nutritive value of three potential complementary foods based on cereal and legumes. *Afr J Food Sci Nutr* 2005;5:1-4.
33. Spackman D, Stein EH, Mooxe S. Automatic recording apparatus for use in the chromatography of amino acids. *Anal Chem* 1958;30:1190-206.
34. Ugwu FM, Ekwu FC. Studies on the Nutritional Quality of Some Diets from Breadfruit (*Treculia africana*). Lagos: Paper Presented at the 20th Annual Conference of the Nigerian Institute of Food Science and Technology. Book of Abstract, 13. 1996.
35. Umezuruike AC, Nwabueze TU, Akobundu EN. Anti nutrients and hemagglutinin activity of roasted African breadfruit seeds produced under extreme conditions. *Fuuast J Biol* 2016;6:135-40.
36. Web MD. Protein: Are You Getting Enough? Available from: <http://WebMd.com>. [Last accessed on 2014 Sep 05].
37. Woolf PJ, Fua LL, Basu A. V protein: Identifying optimal amino-acid complements from plant based foods. *PLoS One* 2011;6:ei8836.
38. Young VR, Bier DM, Cynober L, Yuzo N, Kadowaki M. Assessment of Adequate Intake of Dietary Amino-acids. Nice France: Conference Proceedings. International Council on Amino-acid Science; 2003.