



## Phytochemical Profiling and Fluorescence Analysis of Aerial Parts of *Vitis vinefera* for Modern Health Care Systems

**Anita V. Handore**

Department of Microbiology, HPT Arts and RYK  
Science College, Nashik, MS, India

**Sharad. R. Khandelwal**

Department of Microbiology, HPT Arts and RYK  
Science College, Nashik, MS, India

### ABSTRACT

In the modern health care system, phytochemical rich plants have widespread applications. Since the ancient times, *V. vinefera* has been extensively used in various ayurvedic preparations due to presence of different bioactive compounds. However, its phyto-constituents shows variation on basis of variety and different parts of plant. The aim of this research is to study the phytochemical profiling and fluorescence analysis of aerial parts by use of distinct varieties of *Vitis vinefera*. Healthy aerial parts (leaf lamina, stem and petiole) of different black and white varieties of *Vitis vinefera* were randomly collected. Organic and aqueous extracts were prepared by soaking the shade dried powder (10%) of each part into 70% ethanol and sterile distill water separately. Phytochemical profiling and fluorescent analysis was carried out by use of reported methods with few modifications. Experimental evaluation showed high abundance of various bioactive phytochemicals in both organic as well as aqueous extracts. It was revealed that organic extracts of both varieties gave best results for some polyphenolic groups like flavonoids, ellagic acid, tannins as well as other groups like glycosides, alkaloids, diterpenes, coumarins, fats and oils, etc. Only the test for anthocyanin was found to be negative in white variety. It was found that carbohydrates and reducing sugar, proteins and amino

acids, saponins as well as tannins has given comparatively good results in aqueous extract. Findings of the fluorescence analysis demonstrated that powdered plant parts treated with different solvents and reagents has shown specific shades of green, brown, pink, black and yellow colour in visible and ultraviolet light. Whereas, some samples showed white, yellow and green fluorescence under short UV-light (254 nm) and long UV light (365 nm). Therefore, based on the outcome of this study, it can be concluded that almost all the aerial parts of *V. vinefera* are rich natural source of various bioactive phytochemicals with clinically proved therapeutic potential. Further investigation for isolation, characterization and purification of such phyto constituents from these cost effective natural sources may become basis for their promising application in the modern health care systems.

**Keywords:** *Vitis vinefera*, Chronic diseases, Phytochemicals, Fluorescence, Bioactive, Aerial parts, Therapeutic agent

### I. INTRODUCTION

Ayurveda is the most ancient health care system in India which includes descriptions of various medicinal plants having significant role in modern medicine, not because they continue to be used as crude drug preparations, but because they serve as the

source of important phytochemicals with important applications in modern therapy. In recent years, developed countries are turning to use traditional medicinal systems as these potent phytochemicals shows defensive mechanism of action against number of chronic diseases. It is reported that phytochemicals like saponins, terpenoids, flavonoids, tannins, steroids and alkaloids have shown significant anti-inflammatory activity. Some polycyclic glycosides, tannins, and alkaloids have shown hypoglycemic activities. Terpenoids are known to possess analgesic, immunomodulatory, anti-cancer, anti-inflammatory, anti-viral and antimicrobial properties while, steroids help in reducing cholesterol levels and regulates the immune response [1-6]. Proteins and carbohydrates are the building blocks of life, our body needs proteins for repairing and maintaining itself. Whereas, group of coumarins have been increasingly attracting special interest as phytochemicals due to the underlying contributions in prevention and treatment of different diseases viz. anti-inflammatory, anticoagulant, antibacterial, antifungal, antiviral, anticancer, anti-hypertensive, anti-tubercular, anticonvulsant, anti-adipogenic [7-8]. Cardiac glycosides, are also proved as inhibitors of the plasma membrane  $\text{Na}^+/\text{K}^+$ -ATPase and clinically used for the treatment of heart failure. Some studies have suggested that cardiac glycosides target cancer cells selectively and efficiently [9].

Among all the phytochemicals, polyphenols are clinically proved group of promising phyto-constituent associated with reduced risk of various chronic diseases, including cancer, cardiovascular disease and neurodegenerative disorders. They are present in various plants in the form of different compounds like stilbene (Resveratrol), flavonoid, tannin, ellagic acid, flavonoids etc. of which, resveratrol has emerged as a potent molecule due to its outstanding biological properties. It is reported that plants containing such phytochemicals have been effectively used for more than 2000 years in the traditional medicinal preparations like *Drakshasava* made from *Vitis vinifera*. Resveratrol acts as anti-cancer, cardio protective, anti-diabetic, antioxidant, anti-aging (Life extension), anti-inflammatory, anti-microbial and neuroprotective activity [10]. In *V. vinefera* some type of flavonoids like quercetin can shows antioxidant, anti-allergic, anti-inflammatory, antiviral, anti-proliferative, and anti-carcinogenic activities, in addition to positive impact on mammalian metabolism [11]. Ellagic acid has a

variety of benefits for their anti-mutagenic, antimicrobial and antioxidant properties, and inhibitors of human immunodeficiency virus (HIV). It plays important prevention role in formation of various tumors [12]. These phyto-constituents shows characteristic fluorescence property in visible as well as in the UV light which is one of the important parameter of pharmacognostical evaluation. It is stated that when physical and chemical methods are inadequate, the plant material can be identified from their adulterants on the basis of fluorescence analysis.

In *V. vinefera*, phytochemicals have been proved as therapeutic agents. Their therapeutic efficacy against treatment of different diseases and disorders may vary with individual phytochemical, group of phytochemicals working together or phytochemicals working with other substances. Although *V. vinefera* contains variety of phytochemicals, its composition varies greatly among different varieties. Therefore, aim of this research is to study the phytochemical profiling and fluorescence analysis of aerial parts by use of distinct varieties of *Vitis vinefera* for their promising application in the modern health care systems.

## II. MATERIALS AND METHODS

### Plant Material Collection and Extraction

Healthy aerial plant parts (leaf -lamina, Stem and petiole) of different black and white varieties of *Vitis vinefera* viz. Ganesh -white, Sonaka -black, Jumbo -black, Sharad Seedless- black were randomly collected from the vineyard of Nashik region, Maharashtra India during June 2016. Collected plant parts were cleaned, cut into small pieces and shade dried at room temperature for about fifteen days and grounded to fine powder for extraction. Organic and aqueous extracts were prepared by soaking the dry powder (10%) of each part into 70% Ethanol and sterile distill water respectively. Incubation was carried out at room temperature with gentle shaking for 72 h. The supernatants obtained were used for further analysis.

### Phytochemical Profiling

Identification of various bioactive phyto-constituents in organic and aqueous plant extract was carried out by use of reported methods with few modifications. Results were identified by visual observation of

colour change or by precipitate formation on addition of specific reagents to the test solution. [Table.1]

### Florescence analysis

Dried powder samples (0.5gm) of different aerial plant parts of *V.vinefera* were taken into clean and dried test tubes. To each tube 5ml of different organic solvents like glacial acetic acid, acetone, benzene, chloroform, Dil. Hydrochloric acid, Ethanol, 5% FeCl<sub>3</sub>, Conc. Hydrochloric acid, Methanol, 1N NaOH + Methanol, 1N NaOH, Nitric acid, Petroleum ether, Picric acid, Conc. Sulphuric acid were added separately. Then, the tubes were shaken and allowed to stand for about 30 min. The supernatants were observed in visible light, short UV- light (254 nm) and long UV light (365 nm) for their characteristic colour reaction [13]. Colours were recorded on basis of standard colour chart [Table.2]

### III. RESULTS AND DISCUSSION

Phytochemicals play significant role in defense mechanism by protecting the plant from pathogenic attack, insects, ultraviolet radiations and other environmental stresses. It is clinically proved that phytochemicals has ability to reduce the risk of

different chronic diseases and disorders by their ability to neutralize the free radicals responsible for the onset of these diseases.

During the phytochemical profiling, It was found that both organic as well as aqueous extracts showed presence of various bioactive phyto-constituents such as polyphenolic compounds, flavonoids, anthocyanins, carbohydrate, fats and oils, cardiac glycosides, saponin (Fig.1), tannin, alkaloids, ellagic acid, phytosterols, diterpenes, proteins and amino acids, coumarins etc. in high concentrations.

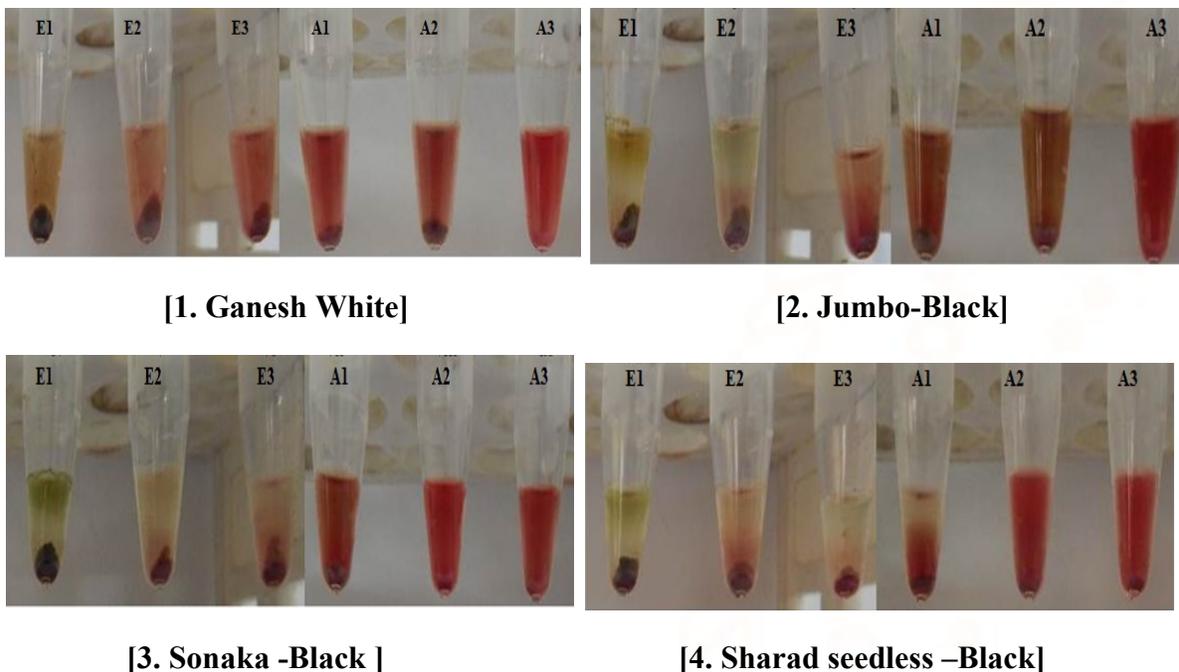
It was revealed that organic extract showed best results for some groups of polyphenols like flavonoids, anthocyanin, ellagic acid as well as other groups like glycosides, alkaloids, diterpenes, fats and oils, coumarins etc. Whereas, test for anthocyanin was negative in white variety. It was found that aqueous extract showed comparatively best results for carbohydrates and reducing sugar, proteins and amino acids, saponins and tannins. Among all the detected compounds, some test for flavonoids, carbohydrates alkaloids, coumarins, protein and acids and tannins showed satisfactory results in both organic as well as aqueous extract. (Table 1)

**Table 1: Phytochemical Screening Tests**

Sr. No.	Group	Test	Reaction	Observation	Reference
1	Phenols	Ferric chloride	0.5 ml extract + 0.5 ml Ferric chloride (10%)	Formation of green precipitate.	[14]
2	Flavonoids	1. Lead acetate	0.5 ml extract + 0.5 ml lead acetate (10%)	Formation of buff coloured solution (pale yellow colour).	[15] [17]
		2. NaOH	0.5 ml extract + 0.5 ml NaOH, (10%) + 0.5 ml dil HCl	Yellow solution turned colourless on addition of dil HCl.	[16-17]
		3. Shinoda	Mg chip (0.5 cm) + 1 ml extract + 2-3 /few drops of conc. HCl	Dark brown solution turns red with effervescence.	[17-18]
		4. Aluminum Chloride	1 ml extract + 0.25 ml aluminum chloride solution (1%), Gentle shaking	Yellow coloration with precipitate	[17] [19]
		5. Sulphuric acid (Anthocyanin)	1.5 ml of extract + few drops of H <sub>2</sub> SO <sub>4</sub>	Development of yellowish orange colour.	[17][20]

3	Carbohydrate	1.Molisch's solution	1 ml of Molisch's solution + 1ml extract + Slowly addition of conc. H <sub>2</sub> SO <sub>4</sub> along the side of test tube.	Appearance of violet ring at the inter phase of the test tube.	[15]
		2.Benedict's reagent	0.5 ml of Benedict's reagent + 0.5 ml extract ,Boil the mixture	Formation of reddish brown colour.	[21]
		3.Reducing sugar (DNSA)	1ml extract + 1ml DNSA ,Boil the mixture	Formation of brown colour.	[22]
4	Cardiac glycoside	Keller-Killiani	2 ml extract + 1.5 ml FeCl <sub>3</sub> (3.5%) + 1ml Glacial acetic acid + Slowly addition of conc. H <sub>2</sub> SO <sub>4</sub> along the side of test tube.	Formation of reddish brown ring at the interphase of tube test	[23]
5	Saponin	Hemolysis	100µl RBC solution (5%) +100 µl extract, Incubation at 37°C,30 Min ,centrifuge 5000rpm for 5 min	Hemolysis observed	[24]
6	Tannin	Gelatin	0.5 ml extract + 1 ml gelatin solution (1 %)	Formation of white precipitate .	[25]
		K <sub>2</sub> Cr <sub>2</sub> O <sub>7</sub>	0.5 ml extract + few drops of K <sub>2</sub> Cr <sub>2</sub> O <sub>7</sub> solution (10 %)	Formation of red precipitate.	[18]
7	Alkaloids	Mayer's reagent	1ml Extract + 1ml Mayer's reagent	Formation of cream colour precipitate .	[20]
8	Ellagic acid	Ellagic acid	1ml Extract + few drops of glacial acetic acid (5% ) (w/v) and + few drops of NaNO <sub>2</sub> solution(5% w/v)	Formation of Muddy or Niger brown precipitate.	[23]
9	Phytosterols	Salkowski's	1ml extract + 1 ml CHCl <sub>3</sub> +Few drops Conc. H <sub>2</sub> SO <sub>4</sub>	Formation of reddish brown colour at interphase of test tube.	[15]
10	Diterpenes	Copper acetate	1 ml Extracts +3-4 drops of copper acetate solution (2%)	Formation of emerald green colour .	[26]
11	Proteins and amino acids	1.Xanthoproteic	1ml extracts + few drops of Conc. Nitric acid	Formation of Yellow colour .	[18]
		2. Ninhydrin	1ml extract + 1ml Ninhydrin reagent (0.25%) , Boiled for few minutes	Formation of blue colour .	[27]
12	Fats and oils	Saponification	1ml Extract+ few drops alcoholic KOH	Formation of soap or partial neutralization	[28]

			(0.5N) + few drops of Phenolphthalein ,heating on a water bath for 1-2 h	of alkali	
13	Coumarins	NaOH	1ml extract + 1.5 ml NaOH (10%)	Formation of yellow colour.	[16]



[E1.Ethanollic Extract -Leaf lamina, E2.Ethanollic Extract -Stem, E3. Ethanollic Extract –Petiole  
A1.Aqueous Extract -Leaf lamina, A2. Aqueous Extract -Stem, A3. Aqueous Extract –Petiole]

**Fig.1. Hemolysis Test for Saponin by Ethanollic and Aqueous Extract of Different Aerial Parts of *V.vinefera***

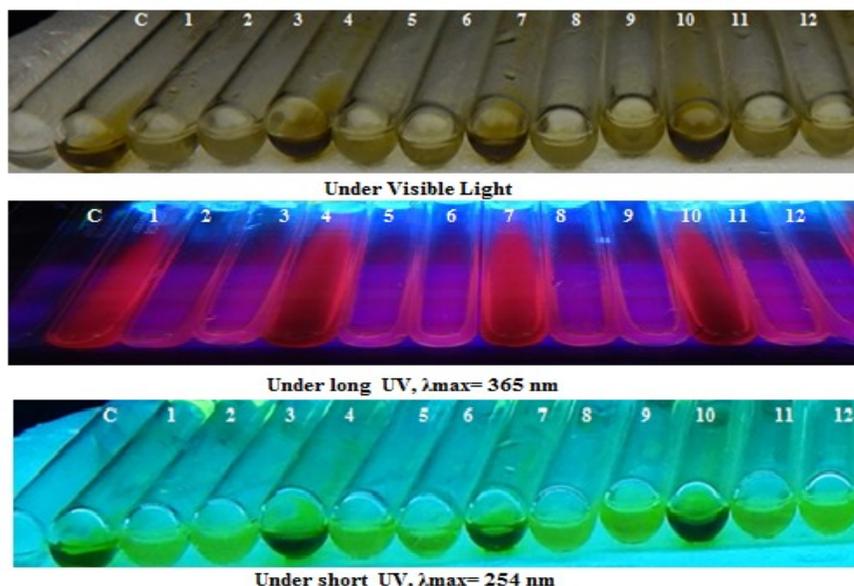
### Fluorescence Analysis

Fluorescence property is exhibited by various chemical constituents present in the plant material. Ultraviolet light produces fluorescence in many natural products which do not shows fluoresce in visible light. Although, some substance are not fluorescent, they may be often converted into fluorescent derivatives by use of different chemical reagents and solvents. Fluorescence is the most important parameter of pharmacognostical evaluation so, qualitative analysis of some crude drugs using fluorescence is widely carried out [29]. There is a

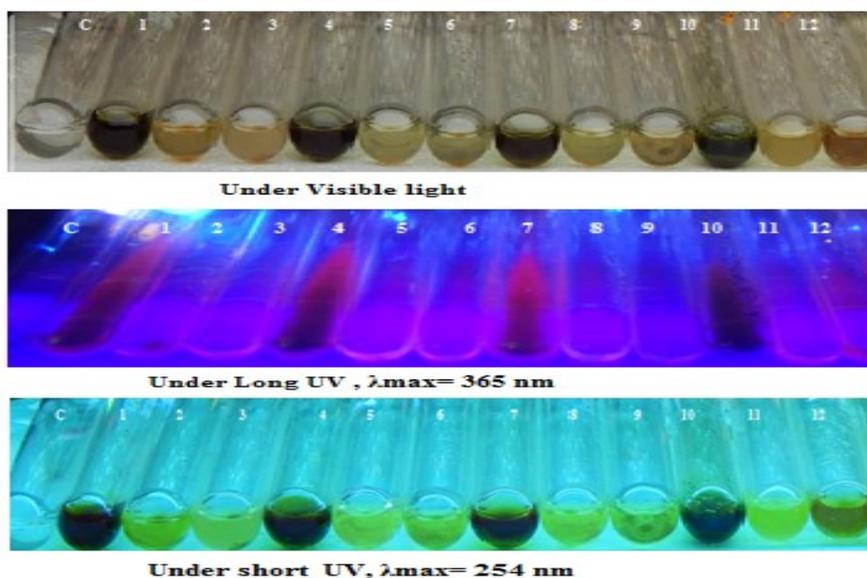
correlation between a compound present in the drugs and their fluorescent behavior at different wavelengths due to the presence of different chemical constituents in the drug. [30]. During the Fluorescence analysis, it was observed that the powdered plant parts of *V.vinefera* showed different shades of green brown, pink and yellow colours in visible and Ultraviolet light ,whereas some of the samples gives white, yellow and green fluorescence under short UV-light (254 nm) and long UV light (365 nm) after treatment with different solvents [Table. 2 and Fig.2.]

**Table.2.Fluorescent analysis of aerial parts of *V.vinefera* in different chemical reagents and solvents under different wavelengths**

Organic Solvents	Visible Light			Under long UV ( $\lambda_{max}$ 365 nm)			Under Short UV ( $\lambda_{max}$ 254 nm)		
	Leaf Lamina	Stem	Petiole	Leaf Lamina	Stem	Petiole	Leaf Lamina	Stem	Petiole
<b>Glacial acetic acid</b>	Deep Olive Green	Brown	Light Brown	Deep Pink	Light Pink	Light Pink	Deep Olive Green	Yellow	Greenish yellow
<b>Acetone</b>	Light Olive Green	Light yellow	Light yellow	Pink	Light Pink	Light Pink	Olive Green	Light yellow	Light yellow
<b>Benzene</b>	Brownish Green	Light Green	light Green	Deep Pink	Pink	Pink	Olive Green	Yellowish green	Yellowish green
<b>Chloroform</b>	Brownish green	Light Brown	Colourless	Deep Pink	Pink	Pink	Olive Green	Yellowish green	Yellowish green
<b>Dil. HCl</b>	Brownish yellow	Light Brown	Light Brown	Pink	Light Pink	Light Pink	Fluorescence Green	Light yellow	Light yellow
<b>Ethanol</b>	Deep Olive Green	Light Olive Green	Light Olive Green	Deep Pink	Light Pink	Light Pink	Deep Olive Green	Fluorescent yellow	Fluorescent yellow
<b>5% Ferric chloride</b>	Deep Brownish green	Greenish orange	Orange	Deep Pink	Light Pink	Light Pink	Deep Olive Green	Deep Olive Green	Light Olive Green
<b>Conc. HCl</b>	Blackish Green	Muddy brown	Muddy brown	Pink	Light Pink	Light Pink	Olive Green	Greenish yellow	Greenish yellow
<b>Methanol</b>	Deep Olive Green	Light Green	Light Green	Deep Pink	Colourless	Colourless	Olive Green	Yellowish green	Yellowish green
<b>1N NaOH +Methanol</b>	Deep Olive Green	Brownish orange	Light brown	Deep Pink	Colourless	Colourless	Deep Olive Green	Deep Olive Green	Light Olive Green
<b>1N NaOH</b>	Deep reddish orange	Reddish orange	Light orange	Deep Pink	Colourless	Colourless	Deep Olive Green	Light Olive Green	Light Olive Green
<b>Nitric Acid</b>	Brownish yellow	Light brownish yellow	Light brownish yellow	Fluorescent white	Fluorescent white	Fluorescent white	Fluorescent Green	Fluorescent Yellow	Fluorescent Yellow
<b>Petroleum Ether</b>	Brownish green	Colourless	Colourless	Pink	Light Pink	Light Pink	Fluorescent Green	Colourless	Colourless
<b>Picric Acid</b>	Greenish orange	Light orange	Light Orange	Blackish pink	Deep pink	Deep pink	Fluorescent Green	Fluorescent Green	Fluorescent Green
<b>Conc. H<sub>2</sub>SO<sub>4</sub></b>	Greenish Brown	Reddish Brown	Reddish Brown	Blackish Pink	Blackish Pink	Blackish Pink	Blackish Green	Blackish Green	Blackish Green



[Fig.2a. Solvent : Benzene]



[Fig.2b. Solvent: Glacial Acetic acid]

C.Negative control, 1.Gaesh –Leaf lamina , 2. Gaesh-Stem, 3.Gaesh –Petiole 4. Jumbo Leaf lamina  
5. Jumbo Stem 6. Jumbo Petiole 7.Sonaka - Leaf lamina 8.Sonaka - Stem 9.Sonaka- Petiole  
10.Sharad Seedless- Leaf lamina , 11. Sharad Seedless- Stem, 12.Sharad Seedless- Petiole

**Fig.2. Fluorescent Analysis of aerial parts of different varieties of *V. vinefera*.**

## CONCLUSION

In the present study, almost all the aerial parts of both varieties of *V. vinefera* showed presence of various phyto-constituents having clinically proved promising role against number of chronic diseases. Results obtained during phytochemical profiling indicate that

both the organic as well as aqueous extracts showed high abundance of various bioactive phytochemicals.

It was revealed that organic solvents showed best results for some polyphenolic groups like flavonoids, anthocyanin, ellagic acid as well as other groups like glycosides, alkaloids, diterpenes, coumarins, fats and oils,etc. whereas, test for anthocyanin was negative in white variety It was found that aqueous extract showed comparatively best results for carbohydrates and reducing sugar, proteins and amino acids, saponins and tannins. It was noticed that some detection test for flavonoids, carbohydrates, alkaloids,

coumarins, protein and acids and tannins gives satisfactory results for both the extracts. Findings of fluorescence analysis demonstrated that some powdered plant parts of *V.vinefera* treated with different solvents and reagents showed specific colour shades viz. green, brown, pink, black and yellow in visible and ultraviolet light. Whereas, some samples showed white, yellow and green fluorescence under short UV- light (254 nm) and long UV light (365 nm). Therefore, based on the outcome of this study, it can be concluded that aerial parts of *V.vinefera* are rich natural source of various bioactive phytochemicals with noteworthy health potential. Further investigation for isolation, characterization and purification of such phyto constituents from these cost effective natural sources may become basis for their promising application in the modern health care systems.

## ACKNOWLEDGEMENT

The authors are grateful to Prin. V. N. Suryavanshi and Dr. L.P.Sharma, HOD, Department of Microbiology, H.P.T .Arts and R.Y.K. Science College, Nashik, India for providing the necessary facilities. We acknowledge J.W.Baviskar and for the assistance. Authors are highly thankful to Mr. D.V.Handore, Research Mentor, Sigma Winery Pvt. Ltd. Nashik for valuable scientific inputs.

## REFERENCES

- 1) H.P Rupasinghe, C.J Jackson, V Poysa, J.Di Berado, J.D. Bewley, J.Jenkinson.,“Soyasapogenol A and B distribution in Soyabean (*Glycine max* (L.) Merr.) in relation to seed physiology ,genetic variability and growing location”, *Journal of Agricultural and Food Chemistry* 51, 2003. pp.5888–5894
- 2) W Salah, NJ Miller, G Pagauga, Tijburg, GP Bolwell, E Rice, C Evans,“Polyphenolic flavonols as scavenger of aqueous phase radicals and chainbreaking antioxidants”,*Arch. Biochem. Biol.* 2, 1995,pp.339- 346
- 3) A. Rio Del ,B G. Obdulio, J.Casfillo, F.G. Main, A.Ortuno,“Uses and properties of citrus flavonoids”, *Afr. J. Biotechnol*,4(7), 1997, pp.685-688.
- 4) K.H Wagner, I Elmadfa, ,“Biological relevance of terpenoids: Overview focusing on mono-di and tetraterpenes”, *Ann. Nutr. Metab*, 47, 2003, pp.95-106.
- 5) B. F Sultana, M.R Anwar, S. Asi.A.S. Chatha, Antioxidant potential of extracts from different agro wastes: Stabilization of corn oil. *Grasas Y Aceites*, 59,2008,pp.205-217
- 6) Rabi , Bishayee A, “Terpenoids and breast cancer chemoprevention. *Breast Cancer Res. Treat.*”, 115,2009,pp.223-239
- 7) K.N Venugopala, V.Rashmi, B. Odhav, “Review on Natural Coumarin Lead Com- pounds for Their Pharmacological Activity”,*BioMed Research International*, 2013,pp.1-14.
- 8) M.E Riveiro, N .De Kimpe, A. Moglioni, R .Vazquez, F. Monczor, C. Shayo, “Coumarins: Old Compounds with Novel Promising Therapeutic Perspectives”,*Curr Med Chem - Anti-Cancer Agents*,17(13), 2010,pp.1325-38.
- 9) M López-Lázaro, “Digi toxin as an anticancer agent with selectivity for cancer cells: possible mechanisms involved”,*Expert Opinion on Therapeutic Targets*, 11,2007,pp.1043-1053
- 10) A.V. Handore and S. R .Khandelwal,“Resveratrol –The Nutraceutical, Whose Real Time Has Come: A Systematic Review”, *International Journal of Advanced Biotechnology and Research*, 8(2), 2017, pp.1516-1544
- 11) W. Ren,Z. Oiao, H. Wang ,L.Zhu and L.Zhang, Flavonoids;Promising anticancer agents *Med.Res.Rev.*,23,2003,pp.519-534
- 12) L. Sepúlveda, A. Ascacio , R.Rodríguez-Herrera, A. Aguilera-Carbó and N Cristóbal.Aguila, , “Ellagic acid:Biological properties and biotechnological development for production processes”,*African Journal of Biotechnology*,10(22), 2011,pp. 4518-4523.
- 13) R. Kavitha, “Fluorescence and Ft-Ir Analysis of Leaf And Fruit of *Trichosanthes Dioica* Roxb”, *World Journal Of Pharmacy And Pharmaceutical Sciences*, 3(10), 2014,pp.563-572
- 14) H Usman, F .I. Abdulrahman, and A Usman, “Qualitative Phytochemical Screening and In Vitro Antimicrobial Effects of Methanol Stem Bark Extract of *Ficus Thoningii* (Moraceae)”, *Afr. J. Trad. CAM*.6 (3), 2009, pp.289 – 295
- 15) S. Mandal, P. Arpita ,A. Samanta, S.Roy, A.Mandal, T.Das Mahapatra, S.Pradhan, K.Das, and D. Kumar Nandi, “Analysis of phytochemical profile of *Terminalia arjuna* bark extract with

- antioxidative and antimicrobial properties”,Asian Pac J Trop Biomed, 3(12),2013,pp.960–966
- 16) M. Majid, Muhammad R. Khan, N. A. Shah, I. Ul Haq, A.F. Muhammad, S. Ullah, A. Sharif, Z. Zahra, T. Younis, and M. Sajid, “Studies on phytochemical, antioxidant, anti-inflammatory and analgesic activities of *Euphorbia dracunculoides*”, BMC Complementary and Alternative Medicine, 2015, pp. 15:349
- 17) A. V. Handore and S. R. Khandelwal, “Screening of *Vitis Vinifera* for flavonoid content and free radical scavenging potential, International Research Journal of Pharmacy, 2017, 8 (8), pp. 1-4
- 18) S. Sabiha., M .Aftab. Ahmad, M. Asif, A Mohd. and S. Ibne, “Physicochemical and phytochemical Standardization of berries of *Myrtus communis* Linn”, J Pharm Bioallied Sci., 4(4), 2012, pp. 322–326
- 19) L. Obasi Nnamdi, C. Egbuonu Anthony. C, O. Ukoha Pius., M Ejikeme Paul, “Comparative phytochemical and antimicrobial screening of some solvent extracts of *Samanea saman* (fabaceae or mimosaceae) pods”, African Journal of Pure and Applied Chemistry, 4(9), 2010, pp. 206-212
- 20) D. Kalita., N Devi., D. Baishya, “Comparative Preliminary Foliar Phytochemical Screening of *Diospyros malabrica* (Desr.) Kostel and *Diospyros lanceifolia* Roxb”, International Journal of Pharmacognosy and Phytochemical Research. 2016; 8(2), pp. 239-243
- 21) C.S. Vimalkumar, V.B Hosagaudar, S.R Suja, V Vilash, N.M Krishnakumar, P.G Latha, “Comparative preliminary phytochemical analysis of ethanolic extracts of leaves of *Olea dioica* Roxb., infected with the rust fungus *Zaghouania oleae* (E.J. Butler) Cummins and non-infected plants”, Journal of Pharmacognosy and Phytochemistry”, 3(4), 2014, pp. 69-72
- 22) V. Alexander, G, Gusakov. E. Kondratyeva, and P Arkady Sinitsyn, “Comparison of Two Methods for Assaying Reducing Sugars in the Determination of Carbohydrase Activities”, International Journal of Analytical Chemistry, 2011, pp. 2-4
- 23) S. Rimjhim., N. Kumari. K, Jainendra, “Preliminary Phytochemical Screening of Methanolic Extract of *Clerodendron infortunatum*”, IOSR Journal of Applied Chemistry, 7, 2014, pp. 110-13
- 24) A Elena. Khatuntseva, M. Vladimir Men’ shov, S Alexander. Shashkov, E. Yury Tsvetkov, Rodion N. Stepanenko, Ya. Raymonda Vlasenko, E Elvira. Shults, A. Genrikh Tolstikov, G Tatjana. Tolstikova, S Dimitri. Baev, A Vasiliy, A Kaledin, Nelli. Popova, Valeriy P. Nikolin, Pavel P. Laktionov, Anna V. Cherepanova, Tatiana V. Kulakovskaya, Ekaterina V. Kulakovskaya and Nikolay E. Nifantiev, “Triterpenoid saponins from the roots of *Acanthophyllum gypsophiloides* Regel”, Beilstein J. Org. Chem. 8, 2012, pp. 763–775.
- 25) S. K. Bhandary, K.N. Suchetha, S. Vadisha Bhat, K.P Sharmila., M. P. Bekal, “Preliminary Phytochemical Screening Of Various Extracts Of *Punica Granatum* Peel, Whole Fruit And Seeds”, Nitte University Journal of Health Science, 2(4), 2012, pp. 2249-7110
- 26) M. Singh, M .Kaur, C.B.S, Dangi, H Singh, “Phytochemical & TLC Profile of *Lawsonia Inermis* (Heena)”, International Journal for Pharmaceutical Research Scholars, 3(1), 2014, pp. 624-634
- 27) G. Iram, M. Sohail, S.A. Muhammad, A.A. Muhammad, “Phytochemical, toxicological and antimicrobial evaluation of *lawsonia inermis* extracts against clinical isolates of pathogenic bacteria, Annals of Clinical Microbiology and Antimicrobials 2013, 12:36
- 28) R. Saad *et al*, “Phytochemical Screening And Antioxidant Activity Of Different Parts From Five Malaysian Herbs”, The Experiment, 2014, Vol. 19(2), pp. 1336-1347
- 29) M.K., Gupta, *et al.*, Pharmacognostical evaluation of *Grewia asiatica* International Journal of Plant Sciences, 2006, 1(2), pp. 249-251.
- 30) S.H Ansari, Essential of Pharmacognosy, 1st Edition Birla publications Pvt. Ltd, New Delhi 2006.